

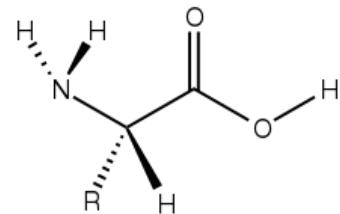
Amino acid

Amino acids are organic compounds that contain amino^[a] ($-\text{NH}_3^+$) and carboxylic acid ($-\text{CO}_2\text{H}$) functional groups, along with a side chain (R group) specific to each amino acid.^[1] The elements present in every amino acid are carbon (C), hydrogen (H), oxygen (O), and nitrogen (N) (CHON); in addition sulfur (S) is present in the side chains of cysteine and methionine, and selenium (Se) in the less common amino acid selenocysteine. More than 500 naturally occurring amino acids are known to constitute monomer units of peptides, including proteins, as of 2020^[2] although only 22 appear in the genetic code, 20 of which have their own designated codons and 2 of which have special coding mechanisms: Selenocysteine which is present in all eukaryotes and pyrrolysine which is present in some prokaryotes.^{[3][4]}

Amino acids are formally named by the IUPAC-IUBMB Joint Commission on Biochemical Nomenclature^[5] in terms of the fictitious "neutral" structure shown in the illustration. For example, the systematic name of alanine is 2-aminopropanoic acid, based on the formula $\text{CH}_3-\text{CH}(\text{NH}_2)-\text{COOH}$. The Commission justified this approach as follows:

The systematic names and formulas given refer to hypothetical forms in which amino groups are unprotonated and carboxyl groups are undissociated. This convention is useful to avoid various nomenclatural problems but should not be taken to imply that these structures represent an appreciable fraction of the amino-acid molecules.

They can be classified according to the locations of the core structural functional groups, as alpha- (α -), beta- (β -), gamma- (γ -) or delta- (δ -) amino acids; other categories relate to polarity, ionization, and side chain group type (aliphatic, acyclic, aromatic, containing hydroxyl or sulfur, etc.). In the form of proteins, amino acid residues form the second-largest component (water being the largest) of human muscles and other tissues.^[6] Beyond their role as residues in proteins, amino acids participate in a number of processes such as neurotransmitter transport and biosynthesis.



Structure of a generic L-amino acid in the "neutral" form needed for defining a systematic name, without implying that this form actually exists in detectable amounts either in aqueous solution or in the solid state.

Contents

History

General structure

- Isomerism
- Side chains
 - Aliphatic side-chains
 - Polar neutral side-chains
 - Amide side-chains
 - Sulfur-containing side-chains
 - Aromatic side-chains
 - Anionic side-chains
 - Cationic side-chains
 - β - and γ -amino acids
- Zwitterions
- Isoelectric point

Physicochemical properties of amino acids

- Table of standard amino acid abbreviations and properties

Occurrence and functions in biochemistry

- Proteinogenic amino acids
- Standard vs nonstandard amino acids
- Non-proteinogenic amino acids
- In human nutrition
- Non-protein functions

Uses in industry

- Expanded genetic code
- Nullomers
- Chemical building blocks
- Biodegradable plastics

Synthesis

[Chemical synthesis](#)

[Biosynthesis](#)

Reactions

[Peptide bond formation](#)

[Catabolism](#)

[Complexation](#)

Chemical analysis

See also

Notes

References

Further reading

External links

History

The first few amino acids were discovered in the early 1800s.^{[7][8]} In 1806, French chemists Louis-Nicolas Vauquelin and Pierre Jean Robiquet isolated a compound from asparagus that was subsequently named asparagine, the first amino acid to be discovered.^{[9][10]} Cystine was discovered in 1810,^[11] although its monomer, cysteine, remained undiscovered until 1884.^{[12][10][b]} Glycine and leucine were discovered in 1820.^[13] The last of the 20 common amino acids to be discovered was threonine in 1935 by William Cumming Rose, who also determined the essential amino acids and established the minimum daily requirements of all amino acids for optimal growth.^{[14][15]}

The unity of the chemical category was recognized by Wurtz in 1865, but he gave no particular name to it.^[16] The first use of the term "amino acid" in the English language dates from 1898,^[17] while the German term, Aminosäure, was used earlier.^[18] Proteins were found to yield amino acids after enzymatic digestion or acid hydrolysis. In 1902, Emil Fischer and Franz Hofmeister independently proposed that proteins are formed from many amino acids, whereby bonds are formed between the amino group of one amino acid with the carboxyl group of another, resulting in a linear structure that Fischer termed "peptide".^[19]

General structure

In the structure shown at the top of the page R represents a side chain specific to each amino acid. The carbon atom next to the carboxyl group is called the α -carbon. Amino acids containing an amino group bonded directly to the α -carbon are referred to as α -amino acids.^[20] These include proline and hydroxyproline,^[c] which are secondary amines. In the past they were often called imino acids, a misnomer because they do not contain an imine grouping $\text{HN}=\text{C}$.^[21] The obsolete term remains frequent.

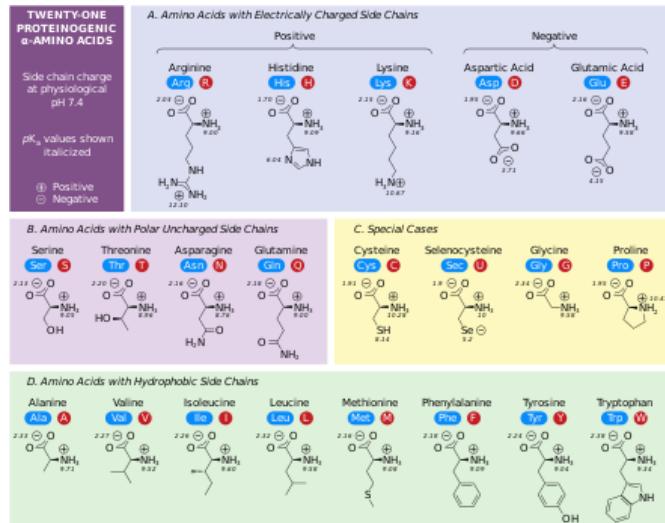
Isomerism

The common natural forms of amino acids have the structure $-\text{NH}_3^+$ ($-\text{NH}_2^-$ in the case of proline) and $-\text{CO}_2^-$ functional groups attached to the same C atom, and are thus α -amino acids. With the exception of achiral glycine, natural amino acids have the L configuration,^[22] and are the only ones found in proteins during translation in the ribosome.

The L and D convention for amino acid configuration refers not to the optical activity of the amino acid itself but rather to the optical activity of the isomer of glyceraldehyde from which that amino acid can, in theory, be synthesized (D-glyceraldehyde is dextrorotatory; L-glyceraldehyde is levorotatory).

An alternative convention is to use the (S) and (R) designators to specify the absolute configuration.^[23] Almost all of the amino acids in proteins are (S) at the α carbon, with cysteine being (R) and glycine non-chiral.^[24] Cysteine has its side chain in the same geometric location as the other amino acids, but the R/S terminology is reversed because sulfur has higher atomic number compared to the carboxyl oxygen which gives the side chain a higher priority by the Cahn-Ingold-Prelog sequence rules, whereas the atoms in most other side chains give them lower priority compared to the carboxyl group.^[23]

D-amino acid residues are found in some proteins, but they are rare.



The 21 proteinogenic α -amino acids found in eukaryotes, grouped according to their side chains' pK_a values and charges carried at physiological pH (7.4)

Side chains

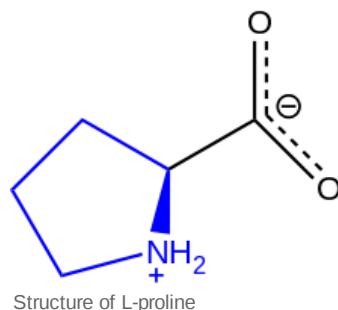
Amino acids are designated as α - when the amino nitrogen atom is attached to the α -carbon, the carbon atom adjacent to the carboxylate group.

In all cases below in this section the pK_a values (if any) refer to the ionization of the groups as amino acid residues in proteins. They are not pK_a values for the free amino acids (which are of little biochemical importance).

Aliphatic side-chains

Several side-chains contain only H and C, and do not ionize. These are as follows (with three- and one-letter symbols in parentheses):

- Glycine (Gly, G): H-
- Alanine (Ala, A): CH₃-
- Valine (Val, V): (CH₃)₂CH-
- Leucine (Leu, L): (CH₃)₂CHCH₂-
- Isoleucine (Ile, I): CH₃CH₂CH(CH₃)
- Proline (Pro, P): -CH₂CH₂CH₂- cyclized onto the amine



Polar neutral side-chains

Two amino acids contain alcohol side-chains. These do not ionize in normal conditions, though one, serine, becomes deprotonated during the catalysis by serine proteases: this is an example of severe perturbation, and is not characteristic of serine residues in general.

- Serine (Ser, S, no pK_a when not severely perturbed): HOCH₂-
- Threonine (Thr, T, no pK_a): CH₃CHOH-

Threonine has two chiral centers, not only the L (2S) chiral center at the α -carbon shared by all amino acids apart from achiral glycine, but also (3R) at the β -carbon. The full stereochemical specification is L-threonine (2S,3R).

Amide side-chains

Two amino acids have amide side-chains, as follows:

- Asparagine (Asn, N): NH₂COCH₂-
- Glutamine (Gln, Q): NH₂COCH₂CH₂-

These side-chains do not ionize in the normal range of pH.

Sulfur-containing side-chains

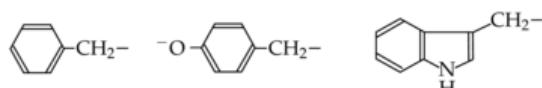
Two side-chains contain sulfur atoms, of which one ionizes in the normal range (with pK_a indicated) and the other does not:

- Cysteine (Cys, C, $pK_a = 8.3$): HSCH₂-
- Methionine (Met, M, no pK_a): CH₃SCH₂CH₂-

Aromatic side-chains

Three amino acids have aromatic ring structures as side-chains, as illustrated. Of these, tyrosine ionizes in the normal range; the other two do not.

- Phenylalanine (Phe, F, no pK_a): left in the illustration
- Tyrosine (Tyr, Y, $pK_a = 9.6$): middle in the illustration
- Tryptophan (Trp, W, no pK_a): right in the illustration



Side-chains of phenylalanine (left), tyrosine (middle) and tryptophan (right)

Anionic side-chains

Two amino acids have side-chains that are anions at ordinary pH. These amino acids are often referred to as if carboxylic acids but are more correctly called carboxylates, as they are deprotonated at most relevant pH values. The anionic carboxylate groups behave as Brønsted bases in all circumstances except for enzymes like pepsin that act in environments of very low pH like the mammalian stomach.

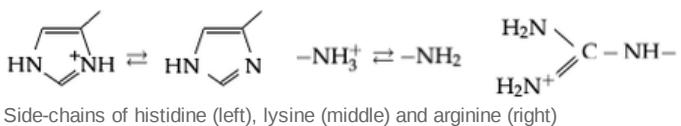
- Aspartate ("aspartic acid", Asp, D, $pK_a = 4.1$): $\text{^O}_2\text{CCH}_2^-$

- Glutamate ("glutamic acid", Glu, E, $pK_a = 4.5$): $\text{O}_2\text{CCH}_2\text{CH}_2^-$

Cationic side-chains

There are three amino acids with side-chains that are cations at neutral pH (though in one, histidine, cationic and neutral forms both exist). They are commonly called *basic amino acids*, but this term is misleading: histidine can act both as a Brønsted acid and as a Brønsted base at neutral pH, lysine acts as a Brønsted acid, and arginine has a fixed positive charge and does not ionize in neutral conditions. The names *histidinium*, *lysinium* and *argininum* would be more accurate names for the structures, but have essentially no currency.

- Histidine (His, H, $pK_a = 6.3$): Protonated and deprotonated forms in equilibrium are shown at the left of the image
- Lysine (Lys, K, $pK_a = 10.4$): Shown in the middle of the image
- Arginine (Arg, R, $pK_a > 12$): Shown at the right of the image



Side-chains of histidine (left), lysine (middle) and arginine (right)

β - and γ -amino acids

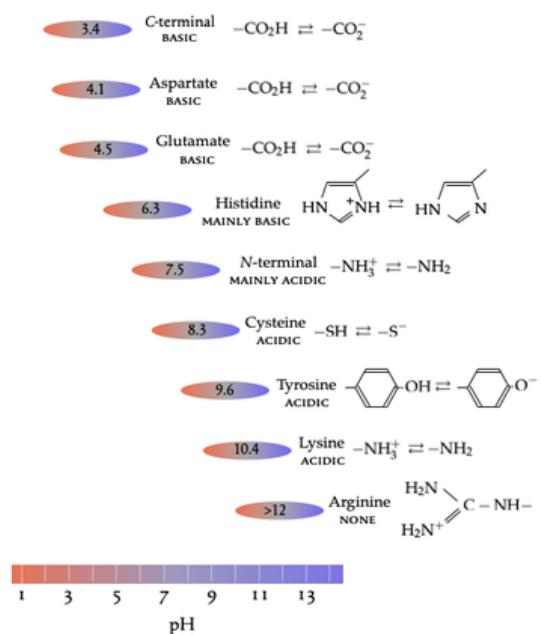
Amino acids with the structure $\text{NH}_3^+-\text{CXY}-\text{CXY}-\text{CO}_2^-$, such as β -alanine, a component of carnosine and a few other peptides, are β -amino acids. Ones with the structure $\text{NH}_3^+-\text{CXY}-\text{CXY}-\text{CXY}-\text{CO}_2^-$ are γ -amino acids, and so on, where X and Y are two substituents (one of which is normally H).^[5]

Zwitterions

In aqueous solution amino acids at moderate pH exist as zwitterions, i.e. as dipolar ions with both NH_3^+ and CO_2^- in charged states, so the overall structure is $\text{NH}_3^+-\text{CHR}-\text{CO}_2^-$. At physiological pH the so-called "neutral forms" $-\text{NH}_2-\text{CHR}-\text{CO}_2\text{H}$ are not present to any measurable degree.^[25] Although the two charges in the real structure add up to zero it is misleading and wrong to call a species with a net charge of zero "uncharged".

At very low pH (below 3), the carboxylate group becomes protonated and the structure becomes an ammonium carboxylic acid, $\text{NH}_3^+-\text{CHR}-\text{CO}_2\text{H}$. This is relevant for enzymes like pepsin that are active in acidic environments such as the mammalian stomach and lysosomes, but does not significantly apply to intracellular enzymes. At very high pH (greater than 10, not normally seen in physiological conditions), the ammonium group is deprotonated to give $\text{NH}_2-\text{CHR}-\text{CO}_2^-$.

Although various definitions of acids and bases are used in chemistry, the only one that is useful for chemistry in aqueous solution is that of Brønsted:^[26] an acid is a species that can donate a proton to another species, and a base is one that can accept a proton. This criterion is used to label the groups in the above illustration. Notice that aspartate and glutamate are the principal groups that act as Brønsted bases, and the common references to these as *acidic amino acids* (together with the C terminal) is completely wrong and misleading. Likewise the so-called *basic amino acids* include one (histidine) that acts as both a Brønsted acid and a base, one (lysine) that acts primarily as a Brønsted acid, and one (arginine) that is normally irrelevant to acid-base behavior as it has a fixed positive charge. In addition, tyrosine and cysteine, which act primarily as acids at neutral pH, are usually forgotten in the usual classification.



Ionization and Brønsted character of N-terminal amino, C-terminal carboxylate, and side chains of amino acid residues

Isoelectric point

For amino acids with uncharged side-chains the zwitterion predominates at pH values between the two pK_a values, but coexists in equilibrium with small amounts of net negative and net positive ions. At the midpoint between the two pK_a values, the trace amount of net negative and trace of net positive ions balance, so that average net charge of all forms present is zero.^[27] This pH is known as the isoelectric point pI , so $pI = \frac{1}{2}(pK_{a1} + pK_{a2})$.

For amino acids with charged side chains, the pK_a of the side chain is involved. Thus for aspartate or glutamate with negative side chains, the terminal amino group is essentially entirely in the charged form NH_3^+ , but this positive charge needs to be balanced by the state with just one C-terminal carboxylate group is negatively charged. This occurs halfway between the two carboxylate pK_a values: $pI = \frac{1}{2}(pK_{a1} + pK_{a(R)})$, where $pK_{a(R)}$ is the side chain pK_a .

Similar considerations apply to other amino acids with ionizable side-chains, including not only glutamate (similar to aspartate), but also cysteine, histidine, lysine, tyrosine and arginine with positive side chains

Amino acids have zero mobility in electrophoresis at their isoelectric point, although this behaviour is more usually exploited for peptides and proteins than single amino acids. Zwitterions have minimum solubility at their isoelectric point, and some amino acids (in particular, with nonpolar side chains) can be isolated by precipitation from water by adjusting the pH to the required isoelectric point.

Physicochemical properties of amino acids

The ca. 20 canonical amino acids can be classified according to their properties. Important factors are charge, hydrophilicity or hydrophobicity, size, and functional groups.^[22] These properties influence protein structure and protein–protein interactions. The water-soluble proteins tend to have their hydrophobic residues (Leu, Ile, Val, Phe, and Trp) buried in the middle of the protein, whereas hydrophilic side chains are exposed to the aqueous solvent. (Note that in biochemistry, a residue refers to a specific monomer within the polymeric chain of a polysaccharide, protein or nucleic acid.) The integral membrane proteins tend to have outer rings of exposed hydrophobic amino acids that anchor them into the lipid bilayer. Some peripheral membrane proteins have a patch of hydrophobic amino acids on their surface that locks onto the membrane. In similar fashion, proteins that have to bind to positively charged molecules have surfaces rich with negatively charged amino acids like glutamate and aspartate, while proteins binding to negatively charged molecules have surfaces rich with positively charged chains like lysine and arginine. For example, lysine and arginine are highly enriched in low-complexity regions of nucleic-acid binding proteins.^[28] There are various hydrophobicity scales of amino acid residues.^[29]

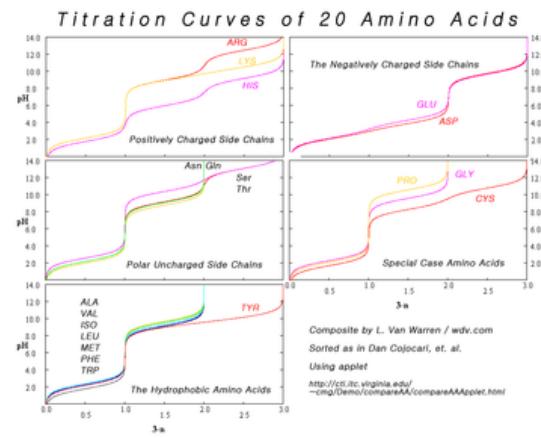
Some amino acids have special properties such as cysteine, that can form covalent disulfide bonds to other cysteine residues, proline that forms a cycle to the polypeptide backbone, and glycine that is more flexible than other amino acids.

Furthermore, glycine and proline are highly enriched within low complexity regions of eukaryotic and prokaryotic proteins, whereas the opposite (under-represented) has been observed for highly reactive, or complex, or hydrophobic amino acids, such as cysteine, phenylalanine, tryptophan, methionine, valine, leucine, isoleucine.^{[28][30][31]}

Many proteins undergo a range of posttranslational modifications, whereby additional chemical groups are attached to the amino acid side chains. Some modifications can produce hydrophobic lipoproteins,^[32] or hydrophilic glycoproteins.^[33] These type of modification allow the reversible targeting of a protein to a membrane. For example, the addition and removal of the fatty acid palmitic acid to cysteine residues in some signaling proteins causes the proteins to attach and then detach from cell membranes.^[34]

Table of standard amino acid abbreviations and properties

Although one-letter symbols are included in the table, IUPAC–IUBMB recommend^[5] that "Use of the one-letter symbols should be restricted to the comparison of long sequences".



Composite of titration curves of twenty proteinogenic amino acids grouped by side chain category

Amino acid	3- and 1-letter symbols		Side chain			Hydropathy index ^[35]	Molar absorptivity ^[36]		Molecular mass	Abundance in proteins (%) ^[37]	Standard genetic coding, IUPAC notation
	3	1	Class	Polarity ^[38]	Net charge at pH 7.4 ^[38]		Wavelength, λ_{\max} (nm)	Coefficient ϵ (mM ⁻¹ ·cm ⁻¹)			
Alanine	Ala	A	Aliphatic	Nonpolar	Neutral	1.8			89.094	8.76	GCN
Arginine	Arg	R	Fixed cation	Basic polar	Positive	-4.5			174.203	5.78	MGR, CGY ^[39]
Asparagine	Asn	N	Amide	Polar	Neutral	-3.5			132.119	3.93	AAY
Aspartate	Asp	D	Anion	Brønsted base	Negative	-3.5			133.104	5.49	GAY
Cysteine	Cys	C	Thiol	Brønsted acid	Neutral	2.5	250	0.3	121.154	1.38	UGY
Glutamine	Gln	Q	Amide	Polar	Neutral	-3.5			146.146	3.9	CAR
Glutamate	Glu	E	Anion	Brønsted base	Negative	-3.5			147.131	6.32	GAR
Glycine	Gly	G	Aliphatic	Nonpolar	Neutral	-0.4			75.067	7.03	GGN
Histidine	His	H	Aromatic cation	Brønsted acid and base	Positive, 10% Neutral, 90%	-3.2	211	5.9	155.156	2.26	CAY
Isoleucine	Ile	I	Aliphatic	Nonpolar	Neutral	4.5			131.175	5.49	AUH
Leucine	Leu	L	Aliphatic	Nonpolar	Neutral	3.8			131.175	9.68	YUR, CUY ^[40]
Lysine	Lys	K	Cation	Brønsted acid	Positive	-3.9			146.189	5.19	AAR
Methionine	Met	M	Thioether	Nonpolar	Neutral	1.9			149.208	2.32	AUG
Phenylalanine	Phe	F	Aromatic	Nonpolar	Neutral	2.8	257, 206, 188	0.2, 9.3, 60.0	165.192	3.87	UUY
Proline	Pro	P	Cyclic	Nonpolar	Neutral	-1.6			115.132	5.02	CCN
Serine	Ser	S	Hydroxylc	Polar	Neutral	-0.8			105.093	7.14	UCN, AGY
Threonine	Thr	T	Hydroxylc	Polar	Neutral	-0.7			119.119	5.53	ACN
Tryptophan	Trp	W	Aromatic	Nonpolar	Neutral	-0.9	280, 219	5.6, 47.0	204.228	1.25	UGG
Tyrosine	Tyr	Y	Aromatic	Brønsted acid	Neutral	-1.3	274, 222, 193	1.4, 8.0, 48.0	181.191	2.91	UAY
Valine	Val	V	Aliphatic	Nonpolar	Neutral	4.2			117.148	6.73	GUN

Two additional amino acids are in some species coded for by codons that are usually interpreted as stop codons:

21st and 22nd amino acids	3-letter	1-letter	Molecular mass
Selenocysteine	Sec	U	168.064
Pyrrolysine	Pyl	O	255.313

In addition to the specific amino acid codes, placeholders are used in cases where chemical or crystallographic analysis of a peptide or protein cannot conclusively determine the identity of a residue. They are also used to summarise conserved protein sequence motifs. The use of single letters to indicate sets of similar residues is similar to the use of abbreviation codes for degenerate bases.^{[41][42]}

Ambiguous amino acids	3-letter	1-letter	Amino acids included	Codons included
Any / unknown	Xaa	X	All	NNN
Asparagine or aspartate	Asx	B	D, N	RAY
Glutamine or glutamate	Glx	Z	E, Q	SAR
Leucine or isoleucine	Xle	J	I, L	YTR, ATH, CTY ^[43]
Hydrophobic		Φ	V, I, L, F, W, Y, M	NTN, TAY, TGG
Aromatic		Ω	F, W, Y, H	YWY, TTY, TGG ^[44]
Aliphatic (non-aromatic)		Ψ	V, I, L, M	VTN, TTR ^[45]
Small		π	P, G, A, S	BCN, RGY, GGR
Hydrophilic		ζ	S, T, H, N, Q, E, D, K, R	VAN, WCN, CGN, AGY ^[46]
Positively-charged		+	K, R, H	ARR, CRY, CGR
Negatively-charged		-	D, E	GAN

Unk is sometimes used instead of Xaa, but is less standard.

Ter or * (from termination) is used in notation for mutations in proteins when a stop codon occurs. It correspond to no amino acid at all.^[47]

In addition, many nonstandard amino acids have a specific code. For example, several peptide drugs, such as Bortezomib and MG132, are artificially synthesized and retain their protecting groups, which have specific codes. Bortezomib is Pyz-Phe-boroLeu, and MG132 is Z-Leu-Leu-Leu-al. To aid in the analysis of protein structure, photo-reactive amino acid analogs are available. These include photoleucine (pLeu) and photomethionine (pMet).^[48]

Occurrence and functions in biochemistry

Amino acids which have the amine group attached to the (alpha-) carbon atom next to the carboxyl group have primary importance in living organisms since they participate in protein synthesis.^[49] They are known as 2-, alpha-, or α-amino acids (generic formula $H_2NCHRCOOH$ in most cases,^[d] where R is an organic substituent known as a "side chain");^[50] often the term "amino acid" is used to refer specifically to these. They include the 22 proteinogenic ("protein-building") amino acids,^{[51][52][53]} which combine into peptide chains ("polypeptides") to form the building blocks of a vast array of proteins.^[49] These are all L-stereoisomers ("left-handed" enantiomers), although a few D-amino acids ("right-handed") occur in bacterial envelopes, as a neuromodulator (D-serine), and in some antibiotics.^[54]

Many proteinogenic and non-proteinogenic amino acids have biological functions. For example, in the human brain, glutamate (standard glutamic acid) and gamma-aminobutyric acid ("GABA", nonstandard gamma-amino acid) are, respectively, the main excitatory and inhibitory neurotransmitters.^[55] Hydroxyproline, a major component of the connective tissue collagen, is synthesised from proline. Glycine is a biosynthetic precursor to porphyrins used in red blood cells. Carnitine is used in lipid transport. Nine proteinogenic amino acids are called "essential" for humans because they cannot be produced from other compounds by the human body and so must be taken in as food. Others may be conditionally essential for certain ages or medical conditions. Essential amino acids may also vary from species to species.^[e] Because of their biological significance, amino acids are important in nutrition and are commonly used in nutritional supplements, fertilizers, feed, and food technology. Industrial uses include the production of drugs, biodegradable plastics, and chiral catalysts.

Proteinogenic amino acids

Amino acids are the precursors to proteins. They join by condensation reactions to form short polymer chains called peptides or longer chains called either polypeptides or proteins. These chains are linear and unbranched, with each amino acid residue within the chain attached to two neighboring amino acids. In Nature, the process of making proteins encoded by DNA/RNA genetic material is called translation and involves the step-by-step addition of amino acids to a growing protein chain by a ribozyme that is called a ribosome.^[56] The order in which the amino acids are added is read through the genetic code from an mRNA template, which is an RNA copy of one of the organism's genes.

Twenty-two amino acids are naturally incorporated into polypeptides and are called proteinogenic or natural amino acids.^[22] Of these, 20 are encoded by the universal genetic code. The remaining 2, selenocysteine and pyrrolysine, are incorporated into proteins by unique synthetic mechanisms. Selenocysteine is incorporated when the mRNA being translated includes a SECIS element, which causes the UGA codon to encode selenocysteine instead of a stop codon.^[57] Pyrrolysine is used by some methanogenic archaea in enzymes that they use to produce methane. It is coded for with the codon UAG, which is normally a stop codon in other organisms.^[58] This UAG codon is followed by a PYLIS downstream sequence.^[59]

Several independent evolutionary studies have suggested that Gly, Ala, Asp, Val, Ser, Pro, Glu, Leu, Thr may belong to a group of amino acids that constituted the early genetic code, whereas Cys, Met, Tyr, Trp, His, Phe may belong to a group of amino acids that constituted later additions of the genetic code.^{[60][61][62]}

Standard vs nonstandard amino acids

The 20 amino acids that are encoded directly by the codons of the universal genetic code are called *standard* or *canonical* amino acids. A modified form of methionine (*N*-formylmethionine) is often incorporated in place of methionine as the initial amino acid of proteins in bacteria, mitochondria and chloroplasts. Other amino acids are called *nonstandard* or *non-canonical*. Most of the nonstandard amino acids are also non-proteinogenic (i.e. they cannot be incorporated into proteins during translation), but two of them are proteinogenic, as they can be incorporated translationally into proteins by exploiting information not encoded in the universal genetic code.

The two nonstandard proteinogenic amino acids are selenocysteine (present in many non-eukaryotes as well as most eukaryotes, but not coded directly by DNA) and pyrrolysine (found only in some archaea and at least one bacterium). The incorporation of these nonstandard amino acids is rare. For example, 25 human proteins include selenocysteine in their primary structure,^[63] and the structurally characterized enzymes (selenoenzymes) employ selenocysteine as the catalytic *moiety* in their active sites.^[64] Pyrrolysine and selenocysteine are encoded via variant codons. For example, selenocysteine is encoded by stop codon and SECIS element.^{[65][66][67]}

N-formylmethionine (which is often the initial amino acid of proteins in bacteria, mitochondria, and chloroplasts) is generally considered as a form of methionine rather than as a separate proteinogenic amino acid. Codon-tRNA combinations not found in nature can also be used to "expand" the genetic code and form novel proteins known as alloproteins incorporating non-proteinogenic amino acids.^{[68][69][70]}

Non-proteinogenic amino acids

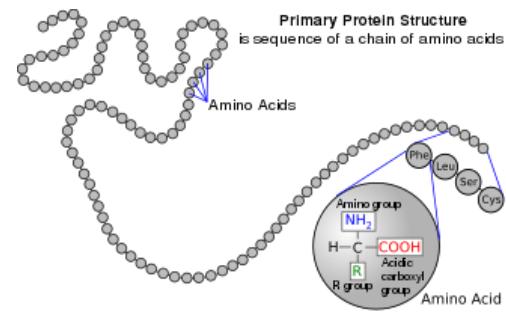
Aside from the 22 proteinogenic amino acids, many *non-proteinogenic* amino acids are known. Those either are not found in proteins (for example carnitine, GABA, levothyroxine) or are not produced directly and in isolation by standard cellular machinery (for example, hydroxyproline and selenomethionine).

Non-proteinogenic amino acids that are found in proteins are formed by post-translational modification, which is modification after translation during protein synthesis. These modifications are often essential for the function or regulation of a protein. For example, the carboxylation of glutamate allows for better binding of calcium cations,^[71] and collagen contains hydroxyproline, generated by hydroxylation of proline.^[72] Another example is the formation of hypusine in the translation initiation factor EIF5A, through modification of a lysine residue.^[73] Such modifications can also determine the localization of the protein, e.g., the addition of long hydrophobic groups can cause a protein to bind to a phospholipid membrane.^[74]

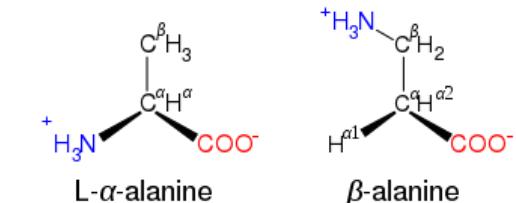
Some non-proteinogenic amino acids are not found in proteins. Examples include 2-aminoisobutyric acid and the neurotransmitter gamma-aminobutyric acid. Non-proteinogenic amino acids often occur as intermediates in the metabolic pathways for standard amino acids – for example, ornithine and citrulline occur in the urea cycle, part of amino acid catabolism (see below).^[75] A rare exception to the dominance of α -amino acids in biology is the β -amino acid beta alanine (3-aminopropanoic acid), which is used in plants and microorganisms in the synthesis of pantothenic acid (vitamin B₅), a component of coenzyme A.^[76]

In human nutrition

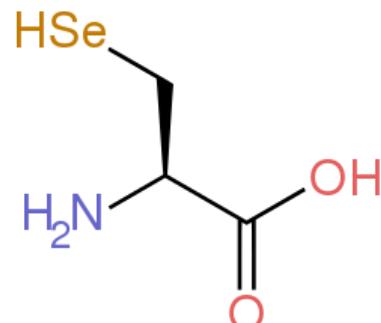
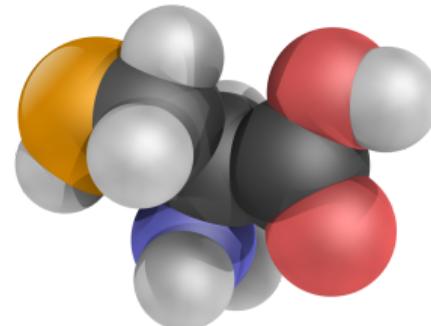
When taken up into the human body from the diet, the 20 standard amino acids either are used to synthesize proteins, other biomolecules, or are oxidized to urea and carbon dioxide as a source of energy.^[77] The oxidation pathway starts with the removal of the amino group by a transaminase; the amino group is then fed into the urea cycle. The other product of transamidation is a keto acid that enters the citric acid cycle.^[78] Glucogenic amino acids can also be converted into glucose, through gluconeogenesis.^[79] Of the 20 standard amino acids, nine (His, Ile, Leu, Lys, Met, Phe, Thr, Trp and Val) are called essential amino acids because the human body cannot synthesize them from other compounds at the level needed for normal growth, so they must be obtained from food.^{[80][81][82]} In addition, cysteine, tyrosine, and arginine are considered semiessential amino acids, and taurine a semiessential aminosulfonic acid in children. The metabolic pathways that synthesize these monomers are not fully developed.^{[83][84]} The amounts required also depend



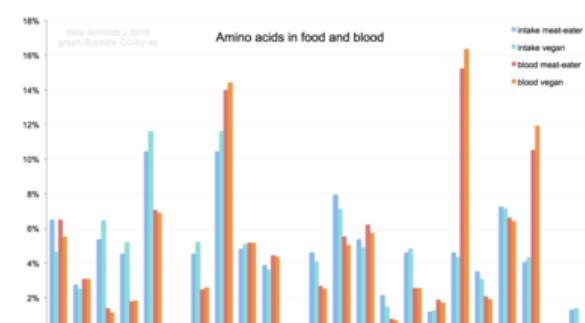
A polypeptide is an unbranched chain of amino acids



β -Alanine and its α -alanine isomer



The amino acid selenocysteine



Share of amino acid in various human diets and the resulting mix of amino acids in human blood serum. Glutamate and glutamine are the most frequent in food at over 10%, while alanine, glutamine, and glycine are the most common in blood.

on the age and health of the individual, so it is hard to make general statements about the dietary requirement for some amino acids. Dietary exposure to the nonstandard amino acid BMAA has been linked to human neurodegenerative diseases, including ALS.^{[85][86]}

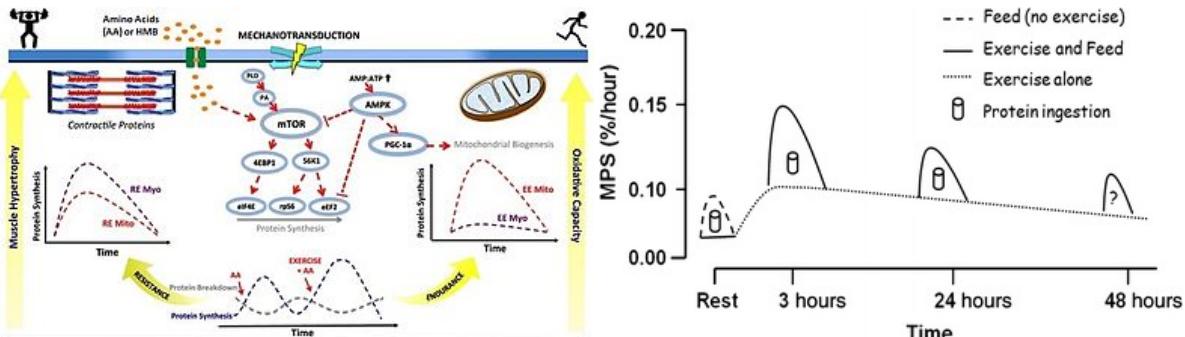


Diagram of the molecular signaling cascades that are involved in myofibrillar muscle protein synthesis and mitochondrial biogenesis in response to physical exercise and specific amino acids or their derivatives (primarily L-leucine and HMB).^[87] Many amino acids derived from food protein promote the activation of mTORC1 and increase protein synthesis by signaling through Rag GTPases.^{[87][88]}

Abbreviations and representations:

- PLD: phospholipase D
- PA: phosphatidic acid
- mTOR: mechanistic target of rapamycin
- AMP: adenosine monophosphate
- ATP: adenosine triphosphate
- AMPK: AMP-activated protein kinase
- PGC-1 α : peroxisome proliferator-activated receptor gamma coactivator-1 α
- S6K1: p70S6 kinase
- 4EBP1: eukaryotic translation initiation factor 4E-binding protein

- eIF4E: eukaryotic translation initiation factor 4E
- RPS6: ribosomal protein S6
- eEF2: eukaryotic elongation factor 2
- RE: resistance exercise; EE: endurance exercise
- Myo: myofibrillar; Mito: mitochondrial
- AA: amino acids
- HMB: β -hydroxy β -methylbutyric acid
- ↑ represents activation
- T represents inhibition

Non-protein functions

In humans, non-protein amino acids also have important roles as metabolic intermediates, such as in the biosynthesis of the neurotransmitter gamma-aminobutyric acid (GABA). Many amino acids are used to synthesize other molecules, for example:

- Tryptophan is a precursor of the neurotransmitter serotonin.^[93]
- Tyrosine (and its precursor phenylalanine) are precursors of the catecholamine neurotransmitters dopamine, epinephrine and norepinephrine and various trace amines.
- Phenylalanine is a precursor of phenethylamine and tyrosine in humans. In plants, it is a precursor of various phenylpropanoids, which are important in plant metabolism.
- Glycine is a precursor of porphyrins such as heme.^[94]

- Arginine is a precursor of nitric oxide.^[95]
- Ornithine and S-adenosylmethionine are precursors of polyamines.^[96]
- Aspartate, glycine, and glutamine are precursors of nucleotides.^[97]
However, not all of the functions of other abundant nonstandard amino acids are known.

Some nonstandard amino acids are used as defenses against herbivores in plants.^[98] For example, canavanine is an analogue of arginine that is found in many legumes,^[99] and in particularly large amounts in *Canavalia gladiata* (sword bean).^[100] This amino acid protects the plants from predators such as insects and can cause illness in people if some types of legumes are eaten without processing.^[101] The non-protein amino acid mimosine is found in other species of legume, in particular *Leucaena leucocephala*.^[102] This compound is an analogue of tyrosine and can poison animals that graze on these plants.

Uses in industry

Amino acids are used for a variety of applications in industry, but their main use is as additives to animal feed. This is necessary, since many of the bulk components of these feeds, such as soybeans, either have low levels or lack some of the essential amino acids: lysine, methionine, threonine, and tryptophan are most important in the production of these feeds.^[103] In this industry, amino acids are also used to chelate metal cations in order to improve the absorption of minerals from supplements, which may be required to improve the health or productivity of these animals.^[104]

The food industry is also a major consumer of amino acids, in particular, glutamic acid, which is used as a flavor enhancer,^[105] and aspartame (aspartylphenylalanine 1-methyl ester) as a low-calorie artificial sweetener.^[106] Similar technology to that used for animal nutrition is employed in the human nutrition industry to alleviate symptoms of mineral deficiencies, such as anemia, by improving mineral absorption and reducing negative side effects from inorganic mineral supplementation.^[107]

The chelating ability of amino acids has been used in fertilizers for agriculture to facilitate the delivery of minerals to plants in order to correct mineral deficiencies, such as iron chlorosis. These fertilizers are also used to prevent deficiencies from occurring and to improve the overall health of the plants.^[108] The remaining production of amino acids is used in the synthesis of drugs and cosmetics.^[103]

Similarly, some amino acids derivatives are used in pharmaceutical industry. They include 5-HTP (5-hydroxytryptophan) used for experimental treatment of depression,^[109] L-DOPA (L-dihydroxyphenylalanine) for Parkinson's treatment,^[110] and eflorenthine drug that inhibits ornithine decarboxylase and used in the treatment of sleeping sickness.^[111]

Expanded genetic code

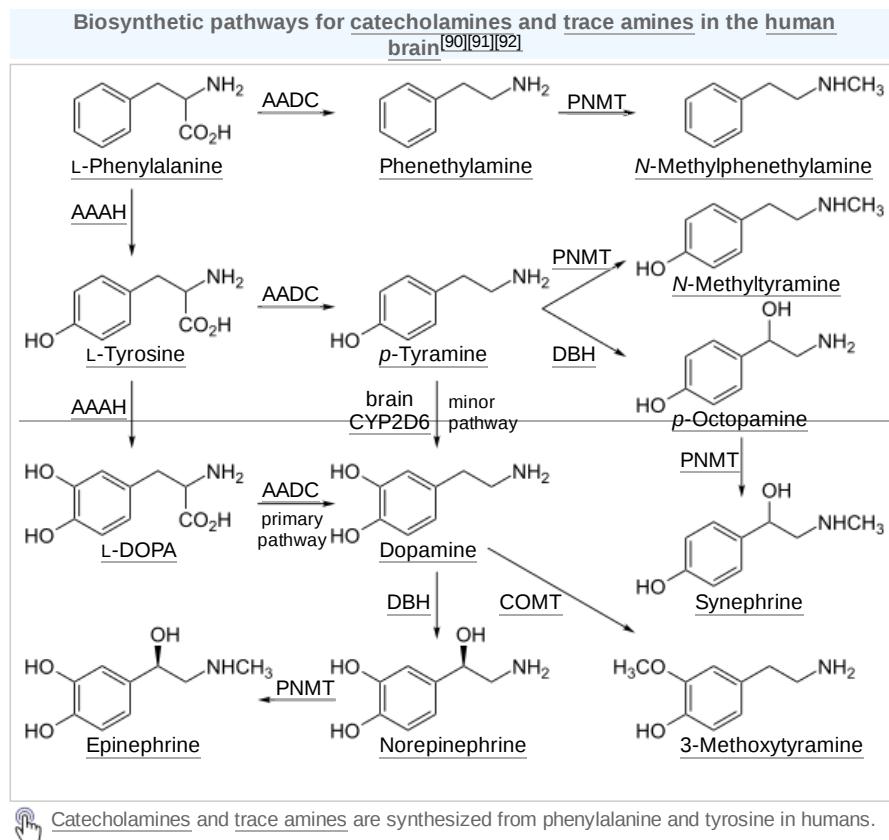
Since 2001, 40 non-natural amino acids have been added into protein by creating a unique codon (recoding) and a corresponding transfer-RNA:aminoacyl – tRNA-synthetase pair to encode it with diverse physicochemical and biological properties in order to be used as a tool to exploring protein structure and function or to create novel or enhanced proteins.^{[68][69]}

Nullomers

Nullomers are codons that in theory code for an amino acid, however, in nature there is a selective bias against using this codon in favor of another, for example bacteria prefer to use CGA instead of AGA to code for arginine.^[112] This creates some sequences that do not appear in the genome. This characteristic can be taken advantage of and used to create new selective cancer-fighting drugs^[113] and to prevent cross-contamination of DNA samples from crime-scene investigations.^[114]

Chemical building blocks

Amino acids are important as low-cost feedstocks. These compounds are used in chiral pool synthesis as enantiomerically pure building blocks.^[115]



Catecholamines and trace amines are synthesized from phenylalanine and tyrosine in humans.

Amino acids have been investigated as precursors chiral catalysts, such as for asymmetric hydrogenation reactions, although no commercial applications exist.^[116]

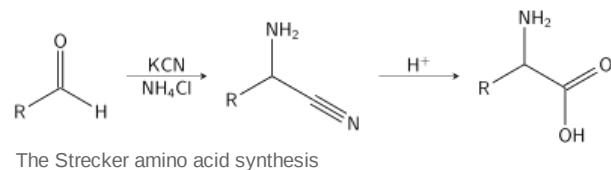
Biodegradable plastics

Amino acids have been considered as components of biodegradable polymers, which have applications as environmentally friendly packaging and in medicine in drug delivery and the construction of prosthetic implants.^[117] An interesting example of such materials is polyaspartate, a water-soluble biodegradable polymer that may have applications in disposable diapers and agriculture.^[118] Due to its solubility and ability to chelate metal ions, polyaspartate is also being used as a biodegradable antiscaling agent and a corrosion inhibitor.^{[119][120]} In addition, the aromatic amino acid tyrosine has been considered as a possible replacement for phenols such as bisphenol A in the manufacture of polycarbonates.^[121]

Synthesis

Chemical synthesis

The commercial production of amino acids usually relies on mutant bacteria that overproduce individual amino acids using glucose as a carbon source. Some amino acids are produced by enzymatic conversions of synthetic intermediates. 2-Aminothiazoline-4-carboxylic acid is an intermediate in one industrial synthesis of L-cysteine for example. Aspartic acid is produced by the addition of ammonia to fumarate using a lyase.^[122]



Biosynthesis

In plants, nitrogen is first assimilated into organic compounds in the form of glutamate, formed from alpha-ketoglutarate and ammonia in the mitochondrion. For other amino acids, plants use transaminases to move the amino group from glutamate to another alpha-keto acid. For example, aspartate aminotransferase converts glutamate and oxaloacetate to alpha-ketoglutarate and aspartate.^[123] Other organisms use transaminases for amino acid synthesis, too.

Nonstandard amino acids are usually formed through modifications to standard amino acids. For example, homocysteine is formed through the transsulfuration pathway or by the demethylation of methionine via the intermediate metabolite S-adenosylmethionine,^[124] while hydroxyproline is made by a post translational modification of proline.^[125]

Microorganisms and plants synthesize many uncommon amino acids. For example, some microbes make 2-aminoisobutyric acid and lanthionine, which is a sulfide-bridged derivative of alanine. Both of these amino acids are found in peptidic lantibiotics such as alamethicin.^[126] However, in plants, 1-aminocyclopropane-1-carboxylic acid is a small disubstituted cyclic amino acid that is an intermediate in the production of the plant hormone ethylene.^[127]

Reactions

Amino acids undergo the reactions expected of the constituent functional groups.^{[128][129]}

Peptide bond formation

As both the amine and carboxylic acid groups of amino acids can react to form amide bonds, one amino acid molecule can react with another and become joined through an amide linkage. This polymerization of amino acids is what creates proteins. This condensation reaction yields the newly formed peptide bond and a molecule of water. In cells, this reaction does not occur directly; instead, the amino acid is first activated by attachment to a transfer RNA molecule through an ester bond. This aminoacyl-tRNA is produced in an ATP-dependent reaction carried out by an aminoacyl tRNA synthetase.^[130] This aminoacyl-tRNA is then a substrate for the ribosome, which catalyzes the attack of the amino group of the elongating protein chain on the ester bond.^[131] As a result of this mechanism, all proteins made by ribosomes are synthesized starting at their *N*-terminus and moving toward their *C*-terminus.

However, not all peptide bonds are formed in this way. In a few cases, peptides are synthesized by specific enzymes. For example, the tripeptide glutathione is an essential part of the defenses of cells against oxidative stress. This peptide is synthesized in two steps from free amino acids.^[132] In the first step, gamma-glutamylcysteine synthetase condenses cysteine and glutamate through a peptide bond formed between the side chain carboxyl of the glutamate (the gamma carbon of this side chain) and the amino group of the cysteine. This dipeptide is then condensed with glycine by glutathione synthetase to form glutathione.^[133]

In chemistry, peptides are synthesized by a variety of reactions. One of the most-used in solid-phase peptide synthesis uses the aromatic oxime derivatives of amino acids as activated units. These are added in sequence onto the growing peptide chain, which is attached to a solid resin support.^[134] Libraries of peptides are used in drug discovery through high-throughput screening.^[135]

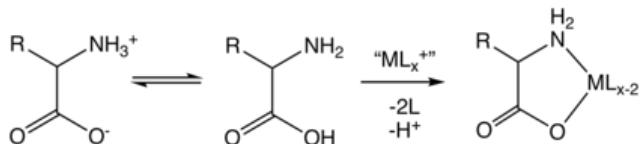
The combination of functional groups allow amino acids to be effective polydentate ligands for metal–amino acid chelates.^[136] The multiple side chains of amino acids can also undergo chemical reactions.

Catabolism

Degradation of an amino acid often involves deamination by moving its amino group to alpha-ketoglutarate, forming glutamate. This process involves transaminases, often the same as those used in amination during synthesis. In many vertebrates, the amino group is then removed through the urea cycle and is excreted in the form of urea. However, amino acid degradation can produce uric acid or ammonia instead. For example, serine dehydratase converts serine to pyruvate and ammonia.^[97] After removal of one or more amino groups, the remainder of the molecule can sometimes be used to synthesize new amino acids, or it can be used for energy by entering glycolysis or the citric acid cycle, as detailed in image at right.

Complexation

Amino acids are bidentate ligands, forming transition metal amino acid complexes.^[138]



Chemical analysis

The total nitrogen content of organic matter is mainly formed by the amino groups in proteins. The Total Kjeldahl Nitrogen (TKN) is a measure of nitrogen widely used in the analysis of (waste) water, soil, food, feed and organic matter in general. As the name suggests, the Kjeldahl method is applied. More sensitive methods are available.^{[139][140]}

See also

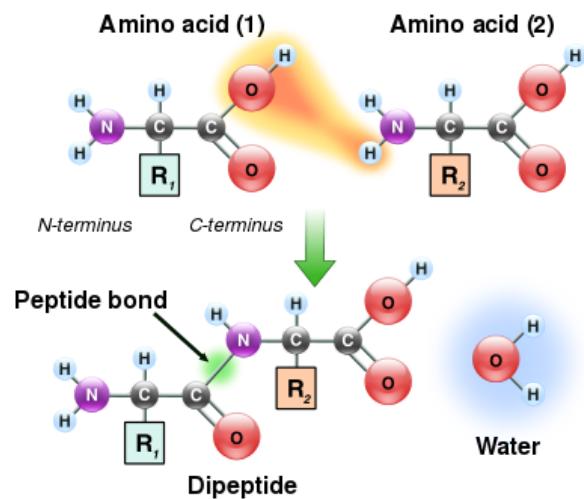
- [Amino acid dating](#)
- [Beta-peptide](#)
- [Degron](#)
- [Erepsin](#)
- [Homochirality](#)
- [Hyperaminoacidemia](#)
- [Leucines](#)
- [Miller–Urey experiment](#)
- [Nucleic acid sequence](#)
- [RNA codon table](#)

Notes

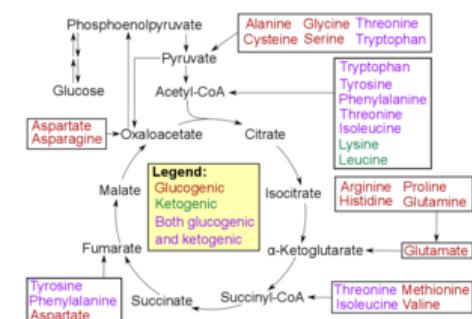
- Strictly *ammonio*, as *amino* refers to uncharged NH_2 , but this term is almost never used.
- The late discovery is explained by the fact that cysteine becomes oxidized to cystine in air.
- Hydroxyproline is present in very few proteins, most notably collagen.
- Proline is an exception to this general formula. It lacks the NH_2 group because of the cyclization of the side chain and is known as an imino acid; it falls under the category of special structured amino acids.
- For example, ruminants such as cows obtain a number of amino acids via microbes in the first two stomach chambers.

References

- Nelson DL, Cox MM (2005). *Principles of Biochemistry* (4th ed.). New York: W. H. Freeman. ISBN 0-7167-4339-6.



The condensation of two amino acids to form a dipeptide. The two amino acid residues are linked through a peptide bond



Catabolism of proteinogenic amino acids. Amino acids can be classified according to the properties of their main degradation products:^[137]

- * *Glucogenic*, with the products having the ability to form glucose by gluconeogenesis
- * *Ketogenic*, with the products not having the ability to form glucose. These products may still be used for ketogenesis or lipid synthesis.
- * Amino acids catabolized into both glucogenic and ketogenic products.

2. Flissi, Areski; Ricart, Emma; Campart, Clémentine; Chevalier, Mickael; Dufresne, Yoann; Michalik, Juraj; Jacques, Philippe; Flahaut, Christophe; Lisacek, Frédérique; Leclère, Valérie; Pupin, Maude (2020). "Norine: update of the nonribosomal peptide resource" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7145658>). *Nucleic Acids Research*. **48** (D1): D465–D469. doi:10.1093/nar/gkz1000 (<https://doi.org/10.1093%2Fnar%2Fgkz1000>). PMC 7145658 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7145658>). PMID 31691799 (<https://pubmed.ncbi.nlm.nih.gov/31691799>).
3. Richard Cammack, ed. (2009). "Newsletter 2009" (<https://web.archive.org/web/20170912194130/http://www.chem.qmul.ac.uk/iubmb/newsletter/2009.html#item35>). Biochemical Nomenclature Committee of IUPAC and NC-IUBMB. Pyrrolysine. Archived from the original (<http://www.chem.qmul.ac.uk/iubmb/newsletter/2009.html#item35>) on 12 September 2017. Retrieved 16 April 2012.
4. Rother, Michael; Krzycki, Joseph A. (1 January 2010). "Selenocysteine, Pyrrolysine, and the Unique Energy Metabolism of Methanogenic Archaea" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2933860>). *Archaea*. **2010**: 1–14. doi:10.1155/2010/453642 (<https://doi.org/10.1155%2F2010%2F453642>). ISSN 1472-3646 (<https://www.worldcat.org/issn/1472-3646>). PMC 2933860 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2933860>). PMID 20847933 (<https://pubmed.ncbi.nlm.nih.gov/20847933>).
5. "Nomenclature and Symbolism for Amino Acids and Peptides" (<https://web.archive.org/web/20081009023202/http://www.chem.qmul.ac.uk/iupac/AminoAcid/AA1n2.html>). IUPAC-IUB Joint Commission on Biochemical Nomenclature. 1983. Archived from the original (<http://www.chem.qmul.ac.uk/iupac/AminoAcid/AA1n2.html>) on 9 October 2008. Retrieved 17 November 2008.
6. Latham MC (1997). "Chapter 8. Body composition, the functions of food, metabolism and energy" (http://www.fao.org/docrep/W0073E/w0073e04.htm#P1625_217364). *Human nutrition in the developing world*. Food and Nutrition Series – No. 29. Rome: Food and Agriculture Organization of the United Nations. Archived (https://web.archive.org/web/20121008212843/http://www.fao.org/docrep/W0073e/w0073e04.htm#P1625_217364) from the original on 8 October 2012. Retrieved 9 September 2012.
7. Vickery HB, Schmidt CL (1931). "The history of the discovery of the amino acids". *Chem. Rev.* **9** (2): 169–318. doi:10.1021/cr60033a001 (<https://doi.org/10.1021%2Fcr60033a001>).
8. Hansen S (May 2015). "Die Entdeckung der proteinogenen Aminosäuren von 1805 in Paris bis 1935 in Illinois" (<https://web.archive.org/web/20171201232937/https://www.arginium.de/wp-content/uploads/2015/12/Entdeckung-der-Aminos%C3%A4uren.pdf>) (PDF) (in German). Berlin. Archived from the original (<https://www.arginium.de/wp-content/uploads/2015/12/Entdeckung-der-Aminos%C3%A4uren.pdf>) (PDF) on 1 December 2017.
9. Vauquelin LN, Robiquet PJ (1806). "The discovery of a new plant principle in Asparagus sativus". *Annales de Chimie*. **57**: 88–93.
10. Anfinsen CB, Edsall JT, Richards FM (1972). *Advances in Protein Chemistry* (<https://archive.org/details/advancesinprotei26anfi/page/99>). New York: Academic Press. pp. 99, 103 (<https://archive.org/details/advancesinprotei26anfi/page/99>). ISBN 978-0-12-034226-6.
11. Wollaston WH (1810). "On cystic oxide, a new species of urinary calculus". *Philosophical Transactions of the Royal Society*. **100**: 223–230. doi:10.1098/rstl.1810.0015 (<https://doi.org/10.1098%2Frstl.1810.0015>). S2CID 110151163 (<https://api.semanticscholar.org/CorpusID:110151163>).
12. Baumann E (1884). "Über cystin und cystein" (<https://web.archive.org/web/20110314075450/http://vlp.mpiwg-berlin.mpg.de/library/data/it16533>). *Z Physiol Chem.* **8** (4): 299–305. Archived from the original (<http://vlp.mpiwg-berlin.mpg.de/library/data/it16533>) on 14 March 2011. Retrieved 28 March 2011.
13. Braconnot HM (1820). "Sur la conversion des matières animales en nouvelles substances par le moyen de l'acide sulfurique". *Annales de Chimie et de Physique*. 2nd Series. **13**: 113–125.
14. Simoni RD, Hill RL, Vaughan M (September 2002). "The discovery of the amino acid threonine: the work of William C. Rose [classical article]" (<http://www.jbc.org/content/277/37/e25>). *The Journal of Biological Chemistry*. **277** (37): E25. doi:10.1016/S0021-9258(20)74369-3 (<https://doi.org/10.1016%2FS0021-9258%2820%2974369-3>). PMID 12218068 (<https://pubmed.ncbi.nlm.nih.gov/12218068>). Archived (<https://web.archive.org/web/20190610150715/http://www.jbc.org/content/277/37/e25>) from the original on 10 June 2019. Retrieved 4 July 2015.
15. McCoy RH, Meyer CE, Rose WC (1935). "Feeding Experiments with Mixtures of Highly Purified Amino Acids. VIII. Isolation and Identification of a New Essential Amino Acid" (<https://doi.org/10.1016%2FS0021-9258%2818%2974986-7>). *Journal of Biological Chemistry*. **112**: 283–302. doi:10.1016/S0021-9258(18)74986-7 (<https://doi.org/10.1016%2FS0021-9258%2818%2974986-7>).
16. Menten, P. *Dictionnaire de chimie: Une approche étymologique et historique*. De Boeck, Bruxelles. link (<https://books.google.com/books?id=NKTKDgAAQBAJ>) Archived (<https://web.archive.org/web/20191228193229/https://books.google.com/books?id=NKTKDgAAQBAJ>) 28 December 2019 at the Wayback Machine.
17. Harper D. "amino-" (<https://www.etymonline.com/word/amino->). *Online Etymology Dictionary*. Archived (<https://web.archive.org/web/20171202102757/https://www.etymonline.com/word/amino->) from the original on 2 December 2017. Retrieved 19 July 2010.
18. Paal C (1894). "Ueber die Einwirkung von Phenyl-i-cyanat auf organische Aminosäuren" (<https://web.archive.org/web/20200725075835/https://zenodo.org/record/1425732>). *Berichte der Deutschen Chemischen Gesellschaft*. **27**: 974–979. doi:10.1002/cber.189402701205 (<https://doi.org/10.1002%2Fcber.189402701205>). Archived from the original (<https://zenodo.org/record/1425732>) on 25 July 2020.
19. Fruton JS (1990). "Chapter 5- Emil Fischer and Franz Hofmeister". *Contrasts in Scientific Style: Research Groups in the Chemical and Biochemical Sciences*. Vol. 191. American Philosophical Society. pp. 163–165. ISBN 978-0-87169-191-0.
20. "Alpha amino acid" (<http://www.merriam-webster.com/medical/alpha-amino%20acid>). *The Merriam-Webster.com Medical Dictionary*. Merriam-Webster Inc. Archived (<https://web.archive.org/web/20150103191856/http://www.merriam-webster.com/medical/alpha-amino%20acid>) from the original on 3 January 2015. Retrieved 3 January 2015..

21. IUPAC. *Compendium of Chemical Terminology*, 2nd ed. (the "Gold Book") (1997). Online corrected version: (2006–) "Imino acids (<https://goldbook.iupac.org/I02959.html>)". doi:[10.1351/goldbook.I02959](https://doi.org/10.1351/goldbook.I02959) (<https://doi.org/10.1351%2Fgoldbook.I02959>). Retrieved 2 April 2012
22. Creighton TH (1993). "Chapter 1" (<https://archive.org/details/proteinsstructur0000crei>). *Proteins: structures and molecular properties*. San Francisco: W. H. Freeman. ISBN 978-0-7167-7030-5.
23. Cahn, R.S.; Ingold, C.K.; Prelog, V. (1966). "Specification of Molecular Chirality". *Angewandte Chemie International Edition*. 5 (4): 385–415. doi:[10.1002/anie.196603851](https://doi.org/10.1002/anie.196603851) (<https://doi.org/10.1002%2Fanie.196603851>).
24. Hatem SM (2006). "Gas chromatographic determination of Amino Acid Enantiomers in tobacco and bottled wines" (<https://web.archive.org/web/20090122104055/http://geb.uni-giessen.de/geb/volltexte/2006/3038/index.html>). University of Giessen. Archived from the original (<http://geb.uni-giessen.de/geb/volltexte/2006/3038/index.html>) on 22 January 2009. Retrieved 17 November 2008.
25. Steinhardt, J.; Reynolds, J. A. (1969). *Multiple equilibria in proteins*. New York: Academic Press. pp. 176–21. ISBN 978-0126654509.
26. Brønsted, J. N. (1923). "Einige Bemerkungen über den Begriff der Säuren und Basen" [Remarks on the concept of acids and bases]. *Recueil des Travaux Chimiques des Pays-Bas*. 42 (8): 718–728. doi:[10.1002/recl.19230420815](https://doi.org/10.1002/recl.19230420815) (<https://doi.org/10.1002%2Frecl.19230420815>).
27. Fennema OR (19 June 1996). *Food Chemistry 3rd Ed.* CRC Press. pp. 327–328. ISBN 978-0-8247-9691-4.
28. Ntountoumi C, Vlastaridis P, Mossialos D, Stathopoulos C, Iliopoulos I, Promponas V, et al. (November 2019). "Low complexity regions in the proteins of prokaryotes perform important functional roles and are highly conserved" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6821194>). *Nucleic Acids Research*. 47 (19): 9998–10009. doi:[10.1093/nar/gkz730](https://doi.org/10.1093/nar/gkz730) (<https://doi.org/10.1093%2Fnar%2Fgkz730>). PMC 6821194 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6821194>). PMID 31504783 (<https://pubmed.ncbi.nlm.nih.gov/31504783>).
29. Urry DW (2004). "The change in Gibbs free energy for hydrophobic association: Derivation and evaluation by means of inverse temperature transitions". *Chemical Physics Letters*. 399 (1–3): 177–183. Bibcode:2004CPL...399..177U (<https://ui.adsabs.harvard.edu/abs/2004CPL...399..177U>). doi:[10.1016/S0009-2614\(04\)01565-9](https://doi.org/10.1016/S0009-2614(04)01565-9) (<https://doi.org/10.1016%2FS0009-2614%2804%2901565-9>).
30. Marcotte EM, Pellegrini M, Yeates TO, Eisenberg D (October 1999). "A census of protein repeats". *Journal of Molecular Biology*. 293 (1): 151–60. doi:[10.1006/jmbi.1999.3136](https://doi.org/10.1006/jmbi.1999.3136) (<https://doi.org/10.1006%2Fjmbi.1999.3136>). PMID 10512723 (<https://pubmed.ncbi.nlm.nih.gov/10512723>).
31. Haerty W, Golding GB (October 2010). Bonen L (ed.). "Low-complexity sequences and single amino acid repeats: not just "junk" peptide sequences". *Genome*. 53 (10): 753–62. doi:[10.1139/G10-063](https://doi.org/10.1139/G10-063) (<https://doi.org/10.1139%2FG10-063>). PMID 20962881 (<https://pubmed.ncbi.nlm.nih.gov/20962881>).
32. Magee T, Seabra MC (April 2005). "Fatty acylation and prenylation of proteins: what's hot in fat". *Current Opinion in Cell Biology*. 17 (2): 190–196. doi:[10.1016/j.ceb.2005.02.003](https://doi.org/10.1016/j.ceb.2005.02.003) (<https://doi.org/10.1016%2Fj.ceb.2005.02.003>). PMID 15780596 (<https://pubmed.ncbi.nlm.nih.gov/15780596>).
33. Pilobello KT, Mahal LK (June 2007). "Deciphering the glycodome: the complexity and analytical challenge of glycomics". *Current Opinion in Chemical Biology*. 11 (3): 300–305. doi:[10.1016/j.cbpa.2007.05.002](https://doi.org/10.1016/j.cbpa.2007.05.002) (<https://doi.org/10.1016%2Fj.cbpa.2007.05.002>). PMID 17500024 (<https://pubmed.ncbi.nlm.nih.gov/17500024>).
34. Smotrys JE, Linder ME (2004). "Palmitoylation of intracellular signaling proteins: regulation and function". *Annual Review of Biochemistry*. 73 (1): 559–587. doi:[10.1146/annurev.biochem.73.011303.073954](https://doi.org/10.1146/annurev.biochem.73.011303.073954) (<https://doi.org/10.1146%2Fannurev.biochem.73.011303.073954>). PMID 15189153 (<https://pubmed.ncbi.nlm.nih.gov/15189153>).
35. Kyte J, Doolittle RF (May 1982). "A simple method for displaying the hydrophobic character of a protein". *Journal of Molecular Biology*. 157 (1): 105–132. CiteSeerX 10.1.1.458.454 (<https://citeseerx.ist.psu.edu/viewdoc/summary?doi=10.1.1.458.454>). doi:[10.1016/0022-2836\(82\)90515-0](https://doi.org/10.1016/0022-2836(82)90515-0) (<https://doi.org/10.1016%2F0022-2836%2882%2990515-0>). PMID 7108955 (<https://pubmed.ncbi.nlm.nih.gov/7108955>).
36. Freifelder D (1983). *Physical Biochemistry* (2nd ed.). W. H. Freeman and Company. ISBN 978-0-7167-1315-9.
37. Kozlowski LP (January 2017). "Proteome-pI: proteome isoelectric point database" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5210655>). *Nucleic Acids Research*. 45 (D1): D1112–D1116. doi:[10.1093/nar/gkw978](https://doi.org/10.1093/nar/gkw978) (<https://doi.org/10.1093%2Fnar%2Fgkw978>). PMC 5210655 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5210655>). PMID 27789699 (<https://pubmed.ncbi.nlm.nih.gov/27789699>).
38. Hausman RE, Cooper GM (2004). *The cell: a molecular approach*. Washington, D.C: ASM Press. p. 51. ISBN 978-0-87893-214-6.
39. Codons can also be expressed by: CGN, AGR
40. codons can also be expressed by: CUN, UUR
41. Aasland R, Abrams C, Ampe C, Ball LJ, Bedford MT, Cesareni G, Gimona M, Hurley JH, Jarchau T, Lehto VP, Lemmon MA, Linding R, Mayer BJ, Nagai M, Sudol M, Walter U, Winder SJ (February 2002). "Normalization of nomenclature for peptide motifs as ligands of modular protein domains". *FEBS Letters*. 513 (1): 141–144. doi:[10.1111/j.1432-1033.1968.tb00350.x](https://doi.org/10.1111/j.1432-1033.1968.tb00350.x) (<https://doi.org/10.1111%2Fj.1432-1033.1968.tb00350.x>). PMID 11911894 (<https://pubmed.ncbi.nlm.nih.gov/11911894>).
42. IUPAC–IUB Commission on Biochemical Nomenclature (1972). "A one-letter notation for amino acid sequences" (<https://doi.org/10.1351%2Fpac197231040639>). *Pure and Applied Chemistry*. 31 (4): 641–645. doi:[10.1351/pac197231040639](https://doi.org/10.1351/pac197231040639) (<https://doi.org/10.1351%2Fpac197231040639>). PMID 5080161 (<https://pubmed.ncbi.nlm.nih.gov/5080161>).
43. Codons can also be expressed by: CTN, ATH, TTR; MTY, YTR, ATA; MTY, HTA, YTG
44. Codons can also be expressed by: TWY, CAY, TGG
45. Codons can also be expressed by: NTR, VTY
46. Codons can also be expressed by: VAN, WCN, MGY, CGP

47. "HGVS: Sequence Variant Nomenclature, Protein Recommendations" (<http://varnomen.hgvs.org/recommendations/protein/variant/substitution/>). Archived (<https://web.archive.org/web/20210924091505/http://varnomen.hgvs.org/recommendations/protein/variant/substitution/>) from the original on 24 September 2021. Retrieved 23 September 2021.
48. Suchanek M, Radzikowska A, Thiele C (April 2005). "Photo-leucine and photo-methionine allow identification of protein–protein interactions in living cells" (<https://doi.org/10.1038%2Fnmeth752>). *Nature Methods*. **2** (4): 261–267. doi:[10.1038/nmeth752](https://doi.org/10.1038%2Fnmeth752) (<https://doi.org/10.1038%2Fnmeth752>). PMID [15782218](#) (<https://pubmed.ncbi.nlm.nih.gov/15782218>).
49. "Chapter 1: Proteins are the Body's Worker Molecules" (<https://web.archive.org/web/20140607084902/https://publications.nigms.nih.gov/structlife/chapter1.html>). *The Structures of Life*. National Institute of General Medical Sciences. 27 October 2011. Archived from the original (<https://publications.nigms.nih.gov/structlife/chapter1.html>) on 7 June 2014. Retrieved 20 May 2008.
50. Clark, Jim (August 2007). "An introduction to amino acids" (<http://www.chemguide.co.uk/organicprops/aminoacids/background.html>). *chemguide*. Archived (<https://web.archive.org/web/20150430051143/http://www.chemguide.co.uk/organicprops/aminoacids/background.html>) from the original on 30 April 2015. Retrieved 4 July 2015.
51. Jakubke H, Sewald N (2008). "Amino acids" (<https://books.google.com/books?id=doe9NwgJTAsC&pg=PA20>). *Peptides from A to Z: A Concise Encyclopedia*. Germany: Wiley-VCH. p. 20. ISBN [9783527621170](#). Archived (<https://web.archive.org/web/20160517144350/https://books.google.com/books?id=doe9NwgJTAsC&pg=PA20>) from the original on 17 May 2016. Retrieved 5 January 2016 – via Google Books.
52. Pollegioni L, Servi S, eds. (2012). *Unnatural Amino Acids: Methods and Protocols*. Methods in Molecular Biology. Vol. 794. Humana Press. p. v. doi:[10.1007/978-1-61779-331-8](https://doi.org/10.1007/978-1-61779-331-8) (<https://doi.org/10.1007/978-1-61779-331-8>). ISBN [978-1-61779-331-8](#). OCLC [756512314](#) (<https://www.worldcat.org/oclc/756512314>). S2CID [3705304](#) (<https://api.semanticscholar.org/CorpusID:3705304>).
53. Hertweck C (October 2011). "Biosynthesis and Charging of Pyrrolysine, the 22nd Genetically Encoded Amino Acid". *Angewandte Chemie International Edition*. **50** (41): 9540–9541. doi:[10.1002/anie.201103769](https://doi.org/10.1002/anie.201103769) (<https://doi.org/10.1002/anie.201103769>). PMID [21796749](#) (<https://pubmed.ncbi.nlm.nih.gov/21796749>). S2CID [5359077](#) (<https://api.semanticscholar.org/CorpusID:5359077>).
54. Michal G, Schomburg D, eds. (2012). *Biochemical Pathways: An Atlas of Biochemistry and Molecular Biology* (2nd ed.). Oxford: Wiley-Blackwell. p. 5. ISBN [978-0-470-14684-2](#).
55. Petroff OA (December 2002). "GABA and glutamate in the human brain". *The Neuroscientist*. **8** (6): 562–573. doi:[10.1177/1073858402238515](https://doi.org/10.1177/1073858402238515) (<https://doi.org/10.1177/1073858402238515>). PMID [12467378](#) (<https://pubmed.ncbi.nlm.nih.gov/12467378>). S2CID [84891972](#) (<https://api.semanticscholar.org/CorpusID:84891972>).
56. Rodnina MV, Beringer M, Wintermeyer W (January 2007). "How ribosomes make peptide bonds". *Trends in Biochemical Sciences*. **32** (1): 20–26. doi:[10.1016/j.tibs.2006.11.007](https://doi.org/10.1016/j.tibs.2006.11.007) (<https://doi.org/10.1016/j.tibs.2006.11.007>). PMID [17157507](#) (<https://pubmed.ncbi.nlm.nih.gov/17157507>).
57. Driscoll DM, Copeland PR (2003). "Mechanism and regulation of selenoprotein synthesis". *Annual Review of Nutrition*. **23** (1): 17–40. doi:[10.1146/annurev.nutr.23.011702.073318](https://doi.org/10.1146/annurev.nutr.23.011702.073318) (<https://doi.org/10.1146/annurev.nutr.23.011702.073318>). PMID [12524431](#) (<https://pubmed.ncbi.nlm.nih.gov/12524431>).
58. Krzycki JA (December 2005). "The direct genetic encoding of pyrrolysine". *Current Opinion in Microbiology*. **8** (6): 706–712. doi:[10.1016/j.mib.2005.10.009](https://doi.org/10.1016/j.mib.2005.10.009) (<https://doi.org/10.1016/j.mib.2005.10.009>). PMID [16256420](#) (<https://pubmed.ncbi.nlm.nih.gov/16256420>).
59. Théobald-Dietrich A, Giegé R, Rudinger-Thirion J (2005). "Evidence for the existence in mRNAs of a hairpin element responsible for ribosome dependent pyrrolysine insertion into proteins". *Biochimie*. **87** (9–10): 813–817. doi:[10.1016/j.biochi.2005.03.006](https://doi.org/10.1016/j.biochi.2005.03.006) (<https://doi.org/10.1016/j.biochi.2005.03.006>). PMID [16164991](#) (<https://pubmed.ncbi.nlm.nih.gov/16164991>).
60. Wong, J. T.-F. (1975). "A Co-Evolution Theory of the Genetic Code" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC432657>). *Proceedings of the National Academy of Sciences*. **72** (5): 1909–1912. Bibcode:1975PNAS...72.1909T (<https://ui.adsabs.harvard.edu/abs/1975PNAS...72.1909T>). doi:[10.1073/pnas.72.5.1909](https://doi.org/10.1073/pnas.72.5.1909) (<https://doi.org/10.1073/pnas.72.5.1909>). PMC [432657](#) (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC432657>). PMID [1057181](#) (<https://pubmed.ncbi.nlm.nih.gov/1057181>).
61. Trifonov EN (December 2000). "Consensus temporal order of amino acids and evolution of the triplet code". *Gene*. **261** (1): 139–151. doi:[10.1016/S0378-1119\(00\)00476-5](https://doi.org/10.1016/S0378-1119(00)00476-5) ([https://doi.org/10.1016/S0378-1119\(00\)00476-5](https://doi.org/10.1016/S0378-1119(00)00476-5)). PMID [11164045](#) (<https://pubmed.ncbi.nlm.nih.gov/11164045>).
62. Higgs PG, Pudritz RE (June 2009). "A thermodynamic basis for prebiotic amino acid synthesis and the nature of the first genetic code". *Astrobiology*. **9** (5): 483–90. arXiv:0904.0402 (<https://arxiv.org/abs/0904.0402>). Bibcode:2009AsBio...9..483H (<https://ui.adsabs.harvard.edu/abs/2009AsBio...9..483H>). doi:[10.1089/ast.2008.0280](https://doi.org/10.1089/ast.2008.0280) (<https://doi.org/10.1089/ast.2008.0280>). PMID [19566427](#) (<https://pubmed.ncbi.nlm.nih.gov/19566427>). S2CID [9039622](#) (<https://api.semanticscholar.org/CorpusID:D9039622>).
63. Kryukov GV, Castellano S, Novoselov SV, Lobanov AV, Zehtab O, Guigó R, Gladyshev VN (May 2003). "Characterization of mammalian selenoproteomes" (<http://digitalcommons.unl.edu/cgi/viewcontent.cgi?article=1072&context=biochemgladyshev>). *Science*. **300** (5624): 1439–1443. Bibcode:2003Sci...300.1439K (<https://ui.adsabs.harvard.edu/abs/2003Sci...300.1439K>). doi:[10.1126/science.1083516](https://doi.org/10.1126/science.1083516) (<https://doi.org/10.1126/science.1083516>). PMID [12775843](#) (<https://pubmed.ncbi.nlm.nih.gov/12775843>). S2CID [10363908](#) (<https://api.semanticscholar.org/CorpusID:10363908>). Archived (<https://web.archive.org/web/20180723073438/http://digitalcommons.unl.edu/cgi/viewcontent.cgi?article=1072&context=biochemgladyshev>) from the original on 23 July 2018. Retrieved 12 June 2019.
64. Gromer S, Urig S, Becker K (January 2004). "The thioredoxin system—from science to clinic". *Medicinal Research Reviews*. **24** (1): 40–89. doi:[10.1002/med.10051](https://doi.org/10.1002/med.10051) (<https://doi.org/10.1002/med.10051>). PMID [14595672](#) (<https://pubmed.ncbi.nlm.nih.gov/14595672>). S2CID [1944741](#) (<https://api.semanticscholar.org/CorpusID:1944741>).

65. Tjong H (2008). *Modeling Electrostatic Contributions to Protein Folding and Binding* (<https://diginole.lib.fsu.edu/islandora/object/fsu%3A175939>) (PhD thesis). Florida State University. p. 1 footnote. Archived (<https://web.archive.org/web/20200128234717/https://diginole.lib.fsu.edu/islandora/object/fsu:175939>) from the original on 28 January 2020. Retrieved 28 January 2020.
66. Stewart L, Burgin AB (2005). Atta-Ur-Rahman, Springer BA, Caldwell GW (eds.). "Whole Gene Synthesis: A Gene-O-Matic Future" (<https://books.google.com/books?id=VoJw6flISSkC&pg=PA299>). *Frontiers in Drug Design and Discovery*. Bentham Science Publishers. 1: 299. doi:10.2174/1574088054583318 (<https://doi.org/10.2174%2F1574088054583318>). ISBN 978-1-60805-199-1. ISSN 1574-0889 (<https://www.worldcat.org/issn/1574-0889>). Archived (<https://web.archive.org/web/20210414240111/https://books.google.com/books?id=VoJw6flISSkC&pg=PA299>) from the original on 14 April 2021. Retrieved 5 January 2016.
67. Elzanowski A, Ostell J (7 April 2008). "The Genetic Codes" (<https://www.ncbi.nlm.nih.gov/Taxonomy/Utils/wprintgc.cgi?mode=c>). National Center for Biotechnology Information (NCBI). Archived (<https://web.archive.org/web/20160820125755/http://130.14.29.110/Taxonomy/Utils/wprintgc.cgi?mode=c>) from the original on 20 August 2016. Retrieved 10 March 2010.
68. Xie J, Schultz PG (December 2005). "Adding amino acids to the genetic repertoire". *Current Opinion in Chemical Biology*. 9 (6): 548–554. doi:10.1016/j.cbpa.2005.10.011 (<https://doi.org/10.1016%2Fj.cbpa.2005.10.011>). PMID 16260173 (<https://pubmed.ncbi.nlm.nih.gov/16260173>).
69. Wang Q, Parrish AR, Wang L (March 2009). "Expanding the genetic code for biological studies" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2696486>). *Chemistry & Biology*. 16 (3): 323–336. doi:10.1016/j.chembiol.2009.03.001 (<https://doi.org/10.1016%2Fj.chembiol.2009.03.001>). PMC 2696486 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2696486>). PMID 19318213 (<https://pubmed.ncbi.nlm.nih.gov/19318213>).
70. Simon M (2005). *Emergent computation: emphasizing bioinformatics* (https://archive.org/details/emergentcomputat00simo_754). New York: AIP Press/Springer Science+Business Media. pp. 105–106 (https://archive.org/details/emergentcomputat00simo_754/page/h116). ISBN 978-0-387-22046-8.
71. Vermeer C (March 1990). "Gamma-carboxyglutamate-containing proteins and the vitamin K-dependent carboxylase" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1131186>). *The Biochemical Journal*. 266 (3): 625–636. doi:10.1042/bj2660625 (<https://doi.org/10.1042%2Fbj2660625>). PMC 1131186 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1131186>). PMID 2183788 (<https://pubmed.ncbi.nlm.nih.gov/2183788>).
72. Bhattacharjee A, Bansal M (March 2005). "Collagen structure: the Madras triple helix and the current scenario". *IUBMB Life*. 57 (3): 161–172. doi:10.1080/15216540500090710 (<https://doi.org/10.1080%2F15216540500090710>). PMID 16036578 (<https://pubmed.ncbi.nlm.nih.gov/16036578>). S2CID 7211864 (<https://api.semanticscholar.org/CorpusID:7211864>).
73. Park MH (February 2006). "The post-translational synthesis of a polyamine-derived amino acid, hypusine, in the eukaryotic translation initiation factor 5A (eIF5A)" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2494880>). *Journal of Biochemistry*. 139 (2): 161–169. doi:10.1093/jb/mvj034 (<https://doi.org/10.1093%2Fjb%2Fmjv034>). PMC 2494880 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2494880>). PMID 16452303 (<https://pubmed.ncbi.nlm.nih.gov/16452303>).
74. Blenis J, Resh MD (December 1993). "Subcellular localization specified by protein acylation and phosphorylation". *Current Opinion in Cell Biology*. 5 (6): 984–989. doi:10.1016/0955-0674(93)90081-Z (<https://doi.org/10.1016%2F0955-0674%2893%2990081-Z>). PMID 8129952 (<https://pubmed.ncbi.nlm.nih.gov/8129952>).
75. Curis E, Nicolis I, Moinard C, Osowska S, Zerrouk N, Bénazeth S, Cynober L (November 2005). "Almost all about citrulline in mammals". *Amino Acids*. 29 (3): 177–205. doi:10.1007/s00726-005-0235-4 (<https://doi.org/10.1007%2Fs00726-005-0235-4>). PMID 16082501 (<https://pubmed.ncbi.nlm.nih.gov/16082501>). S2CID 23877884 (<https://api.semanticscholar.org/CorpusID:23877884>).
76. Coxon KM, Chakauya E, Ottenhof HH, Whitney HM, Blundell TL, Abell C, Smith AG (August 2005). "Pantothenate biosynthesis in higher plants". *Biochemical Society Transactions*. 33 (Pt 4): 743–746. doi:10.1042/BST0330743 (<https://doi.org/10.1042%2FBST0330743>). PMID 16042590 (<https://pubmed.ncbi.nlm.nih.gov/16042590>).
77. Sakami W, Harrington H (1963). "Amino acid metabolism". *Annual Review of Biochemistry*. 32 (1): 355–398. doi:10.1146/annurev.bi.32.070163.002035 (<https://doi.org/10.1146%2Fannurev.bi.32.070163.002035>). PMID 14144484 (<https://pubmed.ncbi.nlm.nih.gov/14144484>).
78. Brosnan JT (April 2000). "Glutamate, at the interface between amino acid and carbohydrate metabolism" (<https://doi.org/10.1093%2Fjn%2F130.4.988S>). *The Journal of Nutrition*. 130 (4S Suppl): 988S–990S. doi:10.1093/jn/130.4.988S (<https://doi.org/10.1093%2Fjn%2F130.4.988S>). PMID 10736367 (<https://pubmed.ncbi.nlm.nih.gov/10736367>).
79. Young VR, Ajami AM (September 2001). "Glutamine: the emperor or his clothes?" (<https://doi.org/10.1093%2Fjn%2F131.9.2449S>). *The Journal of Nutrition*. 131 (9 Suppl): 2449S–2459S, 2486S–2487S. doi:10.1093/jn/131.9.2449S (<https://doi.org/10.1093%2Fjn%2F131.9.2449S>). PMID 11533293 (<https://pubmed.ncbi.nlm.nih.gov/11533293>).
80. Young VR (August 1994). "Adult amino acid requirements: the case for a major revision in current recommendations". *The Journal of Nutrition*. 124 (8 Suppl): 1517S–1523S. doi:10.1093/jn/124.suppl_8.1517S (https://doi.org/10.1093%2Fjn%2F124.suppl_8.1517S). PMID 8064412 (<https://pubmed.ncbi.nlm.nih.gov/8064412>).
81. Fürst P, Stehle P (June 2004). "What are the essential elements needed for the determination of amino acid requirements in humans?" (<https://doi.org/10.1093%2Fjn%2F134.6.1558S>). *The Journal of Nutrition*. 134 (6 Suppl): 1558S–1565S. doi:10.1093/jn/134.6.1558S (<https://doi.org/10.1093%2Fjn%2F134.6.1558S>). PMID 15173430 (<https://pubmed.ncbi.nlm.nih.gov/15173430>).
82. Reeds PJ (July 2000). "Dispensable and indispensable amino acids for humans" (<https://doi.org/10.1093%2Fjn%2F130.7.1835S>). *The Journal of Nutrition*. 130 (7): 1835S–1840S. doi:10.1093/jn/130.7.1835S (<https://doi.org/10.1093%2Fjn%2F130.7.1835S>). PMID 10867060 (<https://pubmed.ncbi.nlm.nih.gov/10867060>).
83. Imura K, Okada A (January 1998). "Amino acid metabolism in pediatric patients". *Nutrition*. 14 (1): 143–148. doi:10.1016/S0899-9007(97)00230-X (<https://doi.org/10.1016%2FS0899-9007%2897%2900230-X>). PMID 9437700 (<https://pubmed.ncbi.nlm.nih.gov/9437700>).

84. Lourenço R, Camilo ME (2002). "Taurine: a conditionally essential amino acid in humans? An overview in health and disease". *Nutricion Hospitalaria*. **17** (6): 262–270. PMID 12514918 (<https://pubmed.ncbi.nlm.nih.gov/12514918>).
85. Holtcamp W (March 2012). "The emerging science of BMAA: do cyanobacteria contribute to neurodegenerative disease?" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3295368>). *Environmental Health Perspectives*. **120** (3): A110–A116. doi:10.1289/ehp.120-a110 (<https://doi.org/10.1289%2Fehp.120-a110>). PMC 3295368 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3295368>). PMID 22382274 (<https://pubmed.ncbi.nlm.nih.gov/22382274>).
86. Cox PA, Davis DA, Mash DC, Metcalf JS, Banack SA (January 2016). "Dietary exposure to an environmental toxin triggers neurofibrillary tangles and amyloid deposits in the brain" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4795023>). *Proceedings Biological Sciences*. **283** (1823): 20152397. doi:10.1098/rspb.2015.2397 (<https://doi.org/10.1098%2Frspb.2015.2397>). PMC 4795023 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4795023>). PMID 26791617 (<https://pubmed.ncbi.nlm.nih.gov/26791617>).
87. Brook MS, Wilkinson DJ, Phillips BE, Perez-Schindler J, Philp A, Smith K, Atherton PJ (January 2016). "Skeletal muscle homeostasis and plasticity in youth and ageing: impact of nutrition and exercise" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4843955>). *Acta Physiologica*. **216** (1): 15–41. doi:10.1111/apha.12532 (<https://doi.org/10.1111%2Fapha.12532>). PMC 4843955 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4843955>). PMID 26010896 (<https://pubmed.ncbi.nlm.nih.gov/26010896>).
88. Lipton JO, Sahn M (October 2014). "The neurology of mTOR" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4223653>). *Neuron*. **84** (2): 275–291. doi:10.1016/j.neuron.2014.09.034 (<https://doi.org/10.1016%2Fj.neuron.2014.09.034>). PMC 4223653 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4223653>). PMID 25374355 (<https://pubmed.ncbi.nlm.nih.gov/25374355>).
- Figure 2: The mTOR Signaling Pathway (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4223653/figure/F2/>) Archived (<https://web.archive.org/web/20200701094741/https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4223653/figure/F2/>) 1 July 2020 at the Wayback Machine
89. Phillips SM (May 2014). "A brief review of critical processes in exercise-induced muscular hypertrophy" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4008813>). *Sports Medicine*. **44** (Suppl. 1): S71–S77. doi:10.1007/s40279-014-0152-3 (<https://doi.org/10.1007%2Fs40279-014-0152-3>). PMC 4008813 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4008813>). PMID 24791918 (<https://pubmed.ncbi.nlm.nih.gov/24791918>).
90. Broadley KJ (March 2010). "The vascular effects of trace amines and amphetamines". *Pharmacology & Therapeutics*. **125** (3): 363–375. doi:10.1016/j.pharmthera.2009.11.005 (<https://doi.org/10.1016%2Fj.pharmthera.2009.11.005>). PMID 19948186 (<https://pubmed.ncbi.nlm.nih.gov/19948186>).
91. Lindemann L, Hoener MC (May 2005). "A renaissance in trace amines inspired by a novel GPCR family". *Trends in Pharmacological Sciences*. **26** (5): 274–281. doi:10.1016/j.tips.2005.03.007 (<https://doi.org/10.1016%2Fj.tips.2005.03.007>). PMID 15860375 (<https://pubmed.ncbi.nlm.nih.gov/15860375>).
92. Wang X, Li J, Dong G, Yue J (February 2014). "The endogenous substrates of brain CYP2D". *European Journal of Pharmacology*. **724**: 211–218. doi:10.1016/j.ejphar.2013.12.025 (<https://doi.org/10.1016%2Fj.ejphar.2013.12.025>). PMID 24374199 (<https://pubmed.ncbi.nlm.nih.gov/24374199>).
93. Savelieva KV, Zhao S, Pogorelov VM, Rajan I, Yang Q, Cullinan E, Lanthorn TH (2008). Bartolomucci A (ed.). "Genetic disruption of both tryptophan hydroxylase genes dramatically reduces serotonin and affects behavior in models sensitive to antidepressants" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2565062>). *PLOS ONE*. **3** (10): e3301. Bibcode:2008PLoS...3.3301S (<https://ui.adsabs.harvard.edu/abs/2008PLoS...3.3301S>). doi:10.1371/journal.pone.0003301 (<https://doi.org/10.1371%2Fjournal.pone.0003301>). PMC 2565062 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2565062>). PMID 18923670 (<https://pubmed.ncbi.nlm.nih.gov/18923670>).
94. Shemin D, Rittenberg D (December 1946). "The biological utilization of glycine for the synthesis of the protoporphyrin of hemoglobin" (<http://www.jbc.org/cgi/reprint/166/2/621>). *The Journal of Biological Chemistry*. **166** (2): 621–625. doi:10.1016/S0021-9258(17)35200-6 (<https://doi.org/10.1016%2FS0021-9258%2817%2935200-6>). PMID 20276176 (<https://pubmed.ncbi.nlm.nih.gov/20276176>). Archived (<https://web.archive.org/web/20220507191150/https://www.jbc.org/cgi/reprint/166/2/621>) from the original on 7 May 2022. Retrieved 3 November 2008.
95. Tejero J, Biswas A, Wang ZQ, Page RC, Haque MM, Hemann C, Zweier JL, Misra S, Stuehr DJ (November 2008). "Stabilization and characterization of a heme-oxy reaction intermediate in inducible nitric-oxide synthase" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2586280>). *The Journal of Biological Chemistry*. **283** (48): 33498–33507. doi:10.1074/jbc.M806122200 (<https://doi.org/10.1074%2Fjbc.M806122200>). PMC 2586280 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2586280>). PMID 18815130 (<https://pubmed.ncbi.nlm.nih.gov/18815130>).
96. Rodríguez-Caso C, Montañez R, Cascante M, Sánchez-Jiménez F, Medina MA (August 2006). "Mathematical modeling of polyamine metabolism in mammals" (<https://doi.org/10.1074%2Fjbc.M602756200>). *The Journal of Biological Chemistry*. **281** (31): 21799–21812. doi:10.1074/jbc.M602756200 (<https://doi.org/10.1074%2Fjbc.M602756200>). PMID 16709566 (<https://pubmed.ncbi.nlm.nih.gov/16709566>).
97. Stryer L, Berg JM, Tymoczko JL (2002). *Biochemistry* (<https://archive.org/details/biochemistry200100jere>) (5th ed.). New York: W.H. Freeman. pp. 693–698 (<https://archive.org/details/biochemistry200100jere/page/693>). ISBN 978-0-7167-4684-3.
98. Hylin JW (1969). "Toxic peptides and amino acids in foods and feeds". *Journal of Agricultural and Food Chemistry*. **17** (3): 492–496. doi:10.1021/jf60163a003 (<https://doi.org/10.1021%2Fjf60163a003>).
99. Turner BL, Harborne JB (1967). "Distribution of canavanine in the plant kingdom". *Phytochemistry*. **6** (6): 863–866. doi:10.1016/S0031-9422(00)86033-1 (<https://doi.org/10.1016%2FS0031-9422%2800%2986033-1>).
100. Ekanayake S, Skog K, Asp NG (May 2007). "Canavanine content in sword beans (Canavalia gladiata): analysis and effect of processing". *Food and Chemical Toxicology*. **45** (5): 797–803. doi:10.1016/j.fct.2006.10.030 (<https://doi.org/10.1016%2Fj.fct.2006.10.030>). PMID 17187914 (<https://pubmed.ncbi.nlm.nih.gov/17187914>).

101. Rosenthal GA (2001). "L-Canavanine: a higher plant insecticidal allelochemical". *Amino Acids*. **21** (3): 319–330. doi:10.1007/s007260170017 (<https://doi.org/10.1007%2Fs007260170017>). PMID 11764412 (<https://pubmed.ncbi.nlm.nih.gov/11764412/>). S2CID 3144019 (<https://api.semanticscholar.org/CorpusID:3144019>).
102. Hammond, Andrew C. (1 May 1995). "Leucaena toxicosis and its control in ruminants" (<https://academic.oup.com/jas/article-abstract/73/5/1487/4718854?redirectedFrom=fulltext>). *Journal of Animal Science*. **73** (5): 1487–1492. doi:10.2527/1995.7351487x (<https://doi.org/10.2527%2F1995.7351487x>). PMID 7665380 (<https://pubmed.ncbi.nlm.nih.gov/7665380/>). Archived (<https://web.archive.org/web/20220507185253/https://academic.oup.com/jas/article-abstract/73/5/1487/4718854?redirectedFrom=fulltext>) from the original on 7 May 2022. Retrieved 7 May 2022.
103. Leuchtenberger W, Huthmacher K, Drauz K (November 2005). "Biotechnological production of amino acids and derivatives: current status and prospects". *Applied Microbiology and Biotechnology*. **69** (1): 1–8. doi:10.1007/s00253-005-0155-y (<https://doi.org/10.1007%2Fs00253-005-0155-y>). PMID 16195792 (<https://pubmed.ncbi.nlm.nih.gov/16195792/>). S2CID 24161808 (<https://api.semanticscholar.org/CorpusID:24161808>).
104. Ashmead HD (1993). *The Role of Amino Acid Chelates in Animal Nutrition*. Westwood: Noyes Publications.
105. Garattini S (April 2000). "Glutamic acid, twenty years later" (<https://doi.org/10.1093%2Fjn%2F130.4.901S>). *The Journal of Nutrition*. **130** (4S Suppl): 901S–909S. doi:10.1093/jn/130.4.901S (<https://doi.org/10.1093%2Fjn%2F130.4.901S>). PMID 10736350 (<https://pubmed.ncbi.nlm.nih.gov/10736350>).
106. Stegink LD (July 1987). "The aspartame story: a model for the clinical testing of a food additive". *The American Journal of Clinical Nutrition*. **46** (1 Suppl): 204–215. doi:10.1093/ajcn/46.1.204 (<https://doi.org/10.1093%2Fajcn%2F46.1.204>). PMID 3300262 (<https://pubmed.ncbi.nlm.nih.gov/3300262>).
107. Albion Laboratories, Inc. "Albion Ferrochel Website" (<http://www.albionferrochel.com>). Archived (<https://web.archive.org/web/20110903054502/http://www.albionferrochel.com/>) from the original on 3 September 2011. Retrieved 12 July 2011.
108. Ashmead HD (1986). *Foliar Feeding of Plants with Amino Acid Chelates*. Park Ridge: Noyes Publications.
109. Turner EH, Loftis JM, Blackwell AD (March 2006). "Serotonin a la carte: supplementation with the serotonin precursor 5-hydroxytryptophan" (<https://escholarship.org/uc/item/58h866d5>). *Pharmacology & Therapeutics*. **109** (3): 325–338. doi:10.1016/j.pharmthera.2005.06.004 (<https://doi.org/10.1016%2Fj.pharmthera.2005.06.004>). PMID 16023217 (<https://pubmed.ncbi.nlm.nih.gov/16023217>). S2CID 2563606 (<https://api.semanticscholar.org/CorpusID:2563606>). Archived (<https://web.archive.org/web/20200413044234/https://escholarship.org/uc/item/58h866d5>) from the original on 13 April 2020. Retrieved 12 June 2019.
110. Kostrzewska RM, Nowak P, Kostrzewska JP, Kostrzewska RA, Brus R (March 2005). "Peculiarities of L-DOPA treatment of Parkinson's disease". *Amino Acids*. **28** (2): 157–164. doi:10.1007/s00726-005-0162-4 (<https://doi.org/10.1007%2Fs00726-005-0162-4>). PMID 15750845 (<https://pubmed.ncbi.nlm.nih.gov/15750845>). S2CID 33603501 (<https://api.semanticscholar.org/CorpusID:33603501>).
111. Heby O, Persson L, Rentala M (August 2007). "Targeting the polyamine biosynthetic enzymes: a promising approach to therapy of African sleeping sickness, Chagas' disease, and leishmaniasis". *Amino Acids*. **33** (2): 359–366. doi:10.1007/s00726-007-0537-9 (<https://doi.org/10.1007%2Fs00726-007-0537-9>). PMID 17610127 (<https://pubmed.ncbi.nlm.nih.gov/17610127>). S2CID 26273053 (<https://api.semanticscholar.org/CorpusID:26273053>).
112. Cruz-Vera LR, Magos-Castro MA, Zamora-Romo E, Guarneros G (2004). "Ribosome stalling and peptidyl-tRNA drop-off during translational delay at AGA codons" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC516057>). *Nucleic Acids Research*. **32** (15): 4462–4468. doi:10.1093/nar/gkh784 (<https://doi.org/10.1093%2Fnar%2Fgkh784>). PMC 516057 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC516057>). PMID 15317870 (<https://pubmed.ncbi.nlm.nih.gov/15317870>).
113. Andy C (October 2012). "Molecules 'too dangerous for nature' kill cancer cells" (<https://www.newscientist.com/article/dn22424-molecules-too-dangerous-for-nature-kill-cancer-cells.html>). *New Scientist*. Archived (<https://web.archive.org/web/20150425171809/http://www.newscientist.com/article/dn22424-molecules-too-dangerous-for-nature-kill-cancer-cells.html>) from the original on 25 April 2015. Retrieved 24 August 2017.
114. "Lethal DNA tags could keep innocent people out of jail" (<https://www.newscientist.com/article/mg21829155.900-lethal-dna-tags-could-keep-innocent-people-out-of-jail.html>). *New Scientist*. 2 May 2013. Archived (<https://web.archive.org/web/20150430163903/http://www.newscientist.com/article/mg21829155.900-lethal-dna-tags-could-keep-innocent-people-out-of-jail.html>) from the original on 30 April 2015. Retrieved 24 August 2017.
115. Hanessian S (1993). "Reflections on the total synthesis of natural products: Art, craft, logic, and the chiron approach". *Pure and Applied Chemistry*. **65** (6): 1189–1204. doi:10.1351/pac199365061189 (<https://doi.org/10.1351%2Fpac199365061189>). S2CID 43992655 (<https://api.semanticscholar.org/CorpusID:43992655>).
116. Blaser HU (1992). "The chiral pool as a source of enantioselective catalysts and auxiliaries". *Chemical Reviews*. **92** (5): 935–952. doi:10.1021/cr00013a009 (<https://doi.org/10.1021%2Fcrr00013a009>).
117. Sanda F, Endo T (1999). "Syntheses and functions of polymers based on amino acids". *Macromolecular Chemistry and Physics*. **200** (12): 2651–2661. doi:10.1002/(SICI)1521-3935(19991201)200:12<2651::AID-MACP2651>3.0.CO;2-P (<https://doi.org/10.1002%2F28SICI%291521-3935%2819991201%29200%3A1%2D3C2651%3A%3AAID-MACP2651%3E3.0.CO%2B2-P>).
118. Gross RA, Kalra B (August 2002). "Biodegradable polymers for the environment" (<https://zenodo.org/record/1231185>). *Science*. **297** (5582): 803–807. Bibcode:2002Sci...297..803G (<https://ui.adsabs.harvard.edu/abs/2002Sci...297..803G>). doi:10.1126/science.297.5582.803 (<https://doi.org/10.1126%2Fscience.297.5582.803>). PMID 12161646 (<https://pubmed.ncbi.nlm.nih.gov/12161646>). Archived (<https://web.archive.org/web/20200725075829/https://zenodo.org/record/1231185>) from the original on 25 July 2020. Retrieved 12 June 2019.
119. Low KC, Wheeler AP, Koskan LP (1996). *Commercial poly(aspartic acid) and Its Uses*. Advances in Chemistry Series. Vol. 248. Washington, D.C.: American Chemical Society.
120. Thombre SM, Sarwade BD (2005). "Synthesis and Biodegradability of Polyaspartic Acid: A Critical Review". *Journal of Macromolecular Science, Part A*. **42** (9): 1299–1315. doi:10.1080/10601320500189604 (<https://doi.org/10.1080%2F10601320500189604>). S2CID 94818855 (<https://api.semanticscholar.org/CorpusID:94818855>).

121. Bourke SL, Kohn J (April 2003). "Polymers derived from the amino acid L-tyrosine: polycarbonates, polyarylates and copolymers with poly(ethylene glycol)". *Advanced Drug Delivery Reviews*. **55** (4): 447–466. doi:10.1016/S0169-409X(03)00038-3 (<https://doi.org/10.1016%2FS0169-409X%2803%2900038-3>). PMID 12706045 (<https://pubmed.ncbi.nlm.nih.gov/12706045>).
122. Drauz K, Grayson I, Kleemann A, Krimmer H, Leuchtenberger W, Weckbecker C (2006). *Ullmann's Encyclopedia of Industrial Chemistry*. Weinheim: Wiley-VCH. doi:10.1002/14356007.a02_057.pub2 (https://doi.org/10.1002%2F14356007.a02_057.pub2).
123. Jones RC, Buchanan BB, Grussem W (2000). *Biochemistry & molecular biology of plants* (<https://archive.org/details/biochemistrymole00buch/page/371>). Rockville, Md: American Society of Plant Physiologists. pp. 371–372 (<https://archive.org/details/biochemistrymole00buch/page/371>). ISBN 978-0-943088-39-6.
124. Brosnan JT, Brosnan ME (June 2006). "The sulfur-containing amino acids: an overview" (<https://doi.org/10.1093%2Fjn%2F136.6.1636S>). *The Journal of Nutrition*. **136** (6 Suppl): 1636S–1640S. doi:10.1093/jn/136.6.1636S (<https://doi.org/10.1093%2Fjn%2F136.6.1636S>). PMID 16702333 (<https://pubmed.ncbi.nlm.nih.gov/16702333>).
125. Kivirikko KI, Pihlajaniemi T (1998). "Collagen hydroxylases and the protein disulfide isomerase subunit of prolyl 4-hydroxylases". *Advances in Enzymology and Related Areas of Molecular Biology*. Advances in Enzymology – and Related Areas of Molecular Biology. Vol. 72. pp. 325–398. doi:10.1002/9780470123188.ch9 (<https://doi.org/10.1002%2F9780470123188.ch9>). ISBN 9780470123188. PMID 9559057 (<https://pubmed.ncbi.nlm.nih.gov/9559057>).
126. Whitmore L, Wallace BA (May 2004). "Analysis of pentaibol sequence composition: implications for in vivo synthesis and channel formation". *European Biophysics Journal*. **33** (3): 233–237. doi:10.1007/s00249-003-0348-1 (<https://doi.org/10.1007%2Fs00249-003-0348-1>). PMID 14534753 (<https://pubmed.ncbi.nlm.nih.gov/14534753>). S2CID 24638475 (<https://api.semanticscholar.org/CorpusID:24638475>).
127. Alexander L, Grierson D (October 2002). "Ethylene biosynthesis and action in tomato: a model for climacteric fruit ripening" (<https://doi.org/10.1093%2Fjxb%2Ferf072>). *Journal of Experimental Botany*. **53** (377): 2039–2055. doi:10.1093/jxb/erf072 (<https://doi.org/10.1093%2Fjxb%2Ferf072>). PMID 12324528 (<https://pubmed.ncbi.nlm.nih.gov/12324528>).
128. Elmore DT, Barrett GC (1998). *Amino acids and peptides* (https://archive.org/details/aminoacidspeptid00barr_040). Cambridge, UK: Cambridge University Press. pp. 48 (https://archive.org/details/aminoacidspeptid00barr_040/page/h64)–60. ISBN 978-0-521-46827-5.
129. Gutteridge A, Thornton JM (November 2005). "Understanding nature's catalytic toolkit". *Trends in Biochemical Sciences*. **30** (11): 622–629. doi:10.1016/j.tibs.2005.09.006 (<https://doi.org/10.1016%2Fj.tibs.2005.09.006>). PMID 16214343 (<https://pubmed.ncbi.nlm.nih.gov/16214343>).
130. Ibba M, Söll D (May 2001). "The renaissance of aminoacyl-tRNA synthesis" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1083889>). *EMBO Reports*. **2** (5): 382–387. doi:10.1093/embo-reports/kve095 (<https://doi.org/10.1093%2Fembo-reports%2Fkve095>). PMC 1083889 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1083889>). PMID 11375928 (<https://pubmed.ncbi.nlm.nih.gov/11375928>).
131. Lengyel P, Söll D (June 1969). "Mechanism of protein biosynthesis" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC378322>). *Bacteriological Reviews*. **33** (2): 264–301. doi:10.1128/MMBR.33.2.264-301.1969 (<https://doi.org/10.1128%2FMMBR.33.2.264-301.1969>). PMC 378322 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC378322>). PMID 4896351 (<https://pubmed.ncbi.nlm.nih.gov/4896351>).
132. Wu G, Fang YZ, Yang S, Lupton JR, Turner ND (March 2004). "Glutathione metabolism and its implications for health" (<https://doi.org/10.1093%2Fjn%2F134.3.489>). *The Journal of Nutrition*. **134** (3): 489–492. doi:10.1093/jn/134.3.489 (<https://doi.org/10.1093%2Fjn%2F134.3.489>). PMID 14988435 (<https://pubmed.ncbi.nlm.nih.gov/14988435>).
133. Meister A (November 1988). "Glutathione metabolism and its selective modification" (<https://doi.org/10.1016%2FS0021-9258%2819%2977815-6>). *The Journal of Biological Chemistry*. **263** (33): 17205–17208. doi:10.1016/S0021-9258(19)77815-6 (<https://doi.org/10.1016%2FS0021-9258%2819%2977815-6>). PMID 3053703 (<https://pubmed.ncbi.nlm.nih.gov/3053703>).
134. Carpin LA (1992). "1-Hydroxy-7-azabenzotriazole. An efficient peptide coupling additive". *Journal of the American Chemical Society*. **115** (10): 4397–4398. doi:10.1021/ja00063a082 (<https://doi.org/10.1021%2Fja00063a082>).
135. Marasco D, Perretta G, Sabatella M, Ruvo M (October 2008). "Past and future perspectives of synthetic peptide libraries". *Current Protein & Peptide Science*. **9** (5): 447–467. doi:10.2174/138920308785915209 (<https://doi.org/10.2174%2F138920308785915209>). PMID 18855697 (<https://pubmed.ncbi.nlm.nih.gov/18855697>).
136. Konara S, Gagnona K, Clearfield A, Thompson C, Hartle J, Ericson C, Nelson C (2010). "Structural determination and characterization of copper and zinc bis-glycinates with X-ray crystallography and mass spectrometry". *Journal of Coordination Chemistry*. **63** (19): 3335–3347. doi:10.1080/00958972.2010.514336 (<https://doi.org/10.1080%2F00958972.2010.514336>). S2CID 94822047 (<https://api.semanticscholar.org/CorpusID:94822047>).
137. Stipanuk MH (2006). *Biochemical, physiological, & molecular aspects of human nutrition* (2nd ed.). Saunders Elsevier.
138. Dghaym RD, Dhawan R, Arndtsen BA (September 2001). "The Use of Carbon Monoxide and Imines as Peptide Derivative Synthons: A Facile Palladium-Catalyzed Synthesis of α-Amino Acid Derived Imidazolines". *Angewandte Chemie*. **40** (17): 3228–3230. doi:10.1002/(SICI)1521-3773(19980703)37:12<1634::AID-ANIE1634>3.0.CO;2-C (<https://doi.org/10.1002%2F%28SICI%291521-3773%2819980703%293A12%3C1634%3A%3AAID-ANIE1634%3E3.0.CO%3B2-C>). PMID 29712039 (<https://pubmed.ncbi.nlm.nih.gov/29712039>).
139. Muñoz-Huerta RF, Guevara-Gonzalez RG, Contreras-Medina LM, Torres-Pacheco I, Prado-Olivarez J, Ocampo-Velazquez RV (August 2013). "A review of methods for sensing the nitrogen status in plants: advantages, disadvantages and recent advances" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3812630>). *Sensors*. Basel, Switzerland. **13** (8): 10823–43. Bibcode:2013Senso..1310823M (<https://ui.adsabs.harvard.edu/abs/2013Senso..1310823M>). doi:10.3390/s130810823 (<https://doi.org/10.3390%2Fs130810823>). PMC 3812630 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3812630>). PMID 23959242 (<https://pubmed.ncbi.nlm.nih.gov/23959242>).

140. Martin PD, Malley DF, Manning G, Fuller L (2002). "Determination of soil organic carbon and nitrogen at the field level using near-infrared spectroscopy". *Canadian Journal of Soil Science*. **82** (4): 413–422. doi:10.4141/S01-054 (<https://doi.org/10.4141/S01-054>).

Further reading

- Tymoczko JL (2012). "Protein Composition and Structure" (<https://archive.org/details/biochemistryseve00berg/page/n61>). *Biochemistry* (<https://archive.org/details/biochemistryseve00berg>). New York: W. H. Freeman and company. pp. 28–31. ISBN 9781429229364.
- Doolittle RF (1989). "Redundancies in protein sequences". In Fasman GD (ed.). *Predictions of Protein Structure and the Principles of Protein Conformation*. New York: Plenum Press. pp. 599–623. ISBN 978-0-306-43131-9. LCCN 89008555 (<http://lccn.loc.gov/89008555>).
- Nelson DL, Cox MM (2000). *Lehninger Principles of Biochemistry* (<https://archive.org/details/lehningerprincip01lehn>) (3rd ed.). Worth Publishers. ISBN 978-1-57259-153-0. LCCN 99049137 (<https://lccn.loc.gov/99049137>).
- Meierhenrich U (2008). *Amino acids and the asymmetry of life* (<https://web.archive.org/web/20120112005425/http://rogov.zwz.ru/Macroevolution/amino.pdf>) (PDF). Berlin: Springer Verlag. ISBN 978-3-540-76885-2. LCCN 2008930865 (<https://lccn.loc.gov/2008930865>). Archived from the original on 12 January 2012.

External links

-  Media related to Amino acid at Wikimedia Commons

Retrieved from "https://en.wikipedia.org/w/index.php?title=Amino_acid&oldid=1099291473"

This page was last edited on 20 July 2022, at 00:39 (UTC).

Text is available under the Creative Commons Attribution-ShareAlike License 3.0; additional terms may apply. By using this site, you agree to the Terms of Use and Privacy Policy. Wikipedia® is a registered trademark of the Wikimedia Foundation, Inc., a non-profit organization.